

FORM PTO-1390  
(REV. 1-98)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

TRANSMITTAL LETTER TO THE UNITED STATES  
DESIGNATED/ELECTED OFFICE (DO/EO/US)  
CONCERNING A FILING UNDER 35 U.S.C. 371

1422-443P

U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

09/673884

INTERNATIONAL APPLICATION NO.

PCT/JP99/02121

INTERNATIONAL FILING DATE

April 21, 1999

PRIORITY DATE CLAIMED

April 23, 1998

TITLE OF INVENTION

METHOD FOR SYNTHESIZING DNA

APPLICANT(S) FOR DO/EO/US

ASADA, Kiyozo; UEMORI, Takashi; SATO, Yoshimi; FUJITA, Tomoko; MIYAKE, Kazue; TAKEDA, Osamu; MUKAI, Hiroyuki; KATO, Ikunoshin

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39 (1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19<sup>th</sup> month from the earliest claimed priority date
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
  - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
  - b. ☒ has been transmitted by the International Bureau.
  - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ A translation of the International Application into English (35 U.S.C. 371(c)(3)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(2)).
  - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
  - b. ☐ have been transmitted by the International Bureau.
  - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
  - d. ☒ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern document(s) or information included:

11. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98./1449-International Search Report with references
12. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A **FIRST** preliminary amendment.  
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information:  
Receipt of Deposit of Microorganisms  
Zero (0) sheets of Formal Drawings  
Sequence Listing (3 pages)

U.S. APPLICATION NO (if known, see 37 CFR 1.5) <div style="font-size: 24pt; font-weight: bold;">09/673884</div>		INTERNATIONAL APPLICATION NO PCT/JP99/02121		ATTORNEY'S DOCKET NUMBER 1422-443P	
--	--	--	--	---------------------------------------	--

17. <input checked="" type="checkbox"/> The following fees are submitted: <b>BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5):</b> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO. .... \$1,000.00  International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO ..... \$860.00  International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO. .... \$710.00  International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) ..... \$690.00  International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4). .... \$100.00 <b>ENTER APPROPRIATE BASIC FEE AMOUNT =</b>	<b>CALCULATIONS      PTO USE ONLY</b>													
	\$	860.00												
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).														
<table border="1" style="width:100%; border-collapse: collapse;"> <tr> <th style="width:20%;">CLAIMS</th> <th style="width:20%;">NUMBER FILED</th> <th style="width:20%;">NUMBER EXTRA</th> <th style="width:20%;">RATE</th> </tr> <tr> <td>Total Claims</td> <td>34 - 20 =</td> <td>14</td> <td>X \$18.00</td> </tr> <tr> <td>Independent Claims</td> <td>4 - 3 =</td> <td>1</td> <td>X \$80.00</td> </tr> </table>	CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	Total Claims	34 - 20 =	14	X \$18.00	Independent Claims	4 - 3 =	1	X \$80.00	\$	252.00
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE											
Total Claims	34 - 20 =	14	X \$18.00											
Independent Claims	4 - 3 =	1	X \$80.00											
MULTIPLE DEPENDENT CLAIM(S) (if applicable) No + \$270.00														
<b>TOTAL OF ABOVE CALCULATIONS =</b>														
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28).														
<b>SUBTOTAL =</b>														
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).														
<b>TOTAL NATIONAL FEE =</b>														
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +														
<b>TOTAL FEES ENCLOSED =</b>														
		Amount to be: \$												
		refunded												
		charged \$												

a. ☒ A check in the amount of \$ 1232.00 to cover the above fees is enclosed.

b. ☐ Please charge my Deposit Account No. \_\_\_\_\_ in the amount of \$ \_\_\_\_\_ to cover the above fees.  
 A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any  
 overpayment to Deposit Account No. 02-2448.

**NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.**

Send all correspondence to:  
**Birch, Stewart, Kolasch & Birch, LLP** or Customer No. 2292  
 P.O. Box 747  
 Falls Church, VA 22040-0747  
 (703)205-8000

SIGNATURE

**WEINER, MARC S.**  
 NAME

**#32.181 (MSW)**  
 REGISTRATION NO.

09/673884

422 Rec'd PCT/PTO 23 OCT 2000

PATENT  
1422-443P

IN THE U.S. PATENT AND TRADEMARK OFFICE

Applicant: ASADA, Kiyozo et al.  
Int'l. Appl. No.: PCT/JP99/02121  
Appl. No.: New Group:  
Filed: October 23, 2000 Examiner:  
For: METHOD FOR SYNTHESIZING DNA

PRELIMINARY AMENDMENT

**BOX PATENT APPLICATION**

Assistant Commissioner for Patents  
Washington, DC 20231

October 23, 2000

Sir:

The following Preliminary Amendments and Remarks are respectfully submitted in connection with the above-identified application.

AMENDMENTS

IN THE SPECIFICATION:

Please amend the specification as follows:

Before line 1, insert --This application is the national phase under 35 U.S.C. § 371 of PCT International Application No. PCT/JP99/02121 which has an International filing date of April 21, 1999, which designated the United States of America.--

IN THE CLAIMS:

CLAIM 4: Line 1, change "claim 2 or 3" to --claim 2--

CLAIM 10: Line 3, change "any one of claims 1 to 9"  
to --claim 9--

CLAIM 15: Line 1, change "any one of claims 10 to 14"  
to --claim 10--

CLAIM 16: Line 2, change "any one of claims 1 to 9"  
to --claim 1--

CLAIM 26: Line 1, change "claim 24 or 25" to --claim 24--

CLAIM 29: Line 1, change "claim 27 or 28" to --claim 27--

CLAIM 30: Line 1, change "any one of claims 27 to 29"  
to --claim 27--

CLAIM 31: Line 2, change "any one of claims 1 to 9" to  
--claim 1--

CLAIM 33: Line 1, change "claim 31 or 32: to --claim 31--

CLAIM 34: Line 1, change "any one of claims 31 to 33"  
to --claim 31--

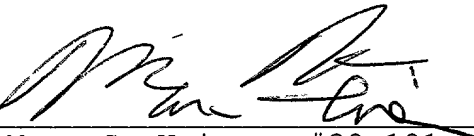
REMARKS

The specification has been amended to provide a cross-reference to the previously filed International Application and to remove the multiple dependencies and places application into better form prior to examination.

If necessary, the Commissioner is hereby authorized in this, concurrent, and future replies, to charge payment or credit any overpayment to Deposit Account No. 02-2448 for any additional fees required under 37 C.F.R. § 1.16 or under 37 C.F.R. § 1.17; particularly, extension of time fees.

Respectfully submitted,

BIRCH, STEWART, KOLASCH & BIRCH, LLP

By   
Marc S. Weiner, #32,181

MSW/dll  
1422-443P

P.O. Box 747  
Falls Church, VA 22040-0747  
(703) 205-8000

(Rev. 04/19/2000)

DESCRIPTIONMETHOD FOR SYNTHESIZING DNA5 TECHNICAL FIELD

The present invention relates to a DNA synthesis reaction-enhancer, a DNA synthesis method, a DNA synthesis reaction composition and a kit usable for the DNA synthesis method, which are useful in the field of genetic engineering.

10

BACKGROUND ART

15

The DNA synthesis is employed for various purposes in the research of the field of genetic engineering. Almost all of them, except for synthesis of short chain DNA such as oligonucleotides, are carried out by the enzymatic method utilizing DNA polymerase. Accordingly, the DNA polymerase is highly valuable for reagents for DNA sequencing, DNA labelling, or site-directed mutagenesis. In addition, recently, thermostable DNA polymerases have been remarked with the developments of the polymerase chain reaction (PCR) method and the reverse transcription-PCR (RT-PCR) in which the PCR method and the reverse transcriptase reaction are combined. Therefore, various kinds of DNA polymerases suitable for PCR method mentioned above have been developed, and commercialized.

20

25

The presently known DNA polymerases can be roughly classified by amino acid sequence homology into four families, among which Family A (pol I type enzymes) and Family B ( $\alpha$ -type enzymes) account for the great majority.

Although the DNA polymerases belonging to the respective families possess generally similar biochemical properties, detailed comparison reveals that depending upon individual enzymes, each of the DNA polymerases has different properties for substrate specificity; substrate analog-incorporating efficiency; degree and rate for primer extension; mode of DNA synthesis; presence or absence of exonuclease activity; optimum reaction conditions such as temperature and pH, and sensitivity against inhibitors. Therefore, the enzyme best suited for the application has so far been selected from the available DNA polymerases.

For example, *Pyrococcus furiosus*, a hyperthermophilic archaebacterium, produces DNA polymerase belonging to  $\alpha$ -type, and the gene thereof has been isolated [*Nucleic Acids Research* 21, 259-265 (1993)]. Recently, novel DNA polymerase showing no structural similarity to any known DNA polymerase was found in the above bacterial strain. In this DNA polymerase, two novel proteins form a complex, whereby exhibiting DNA polymerase activity. In addition, the novel DNA polymerase exhibits potent 3'→5' exonuclease activity and excellent primer extension activity. For example, when the enzyme is used for PCR, a DNA fragment of a size of about 20 kb can be amplified.

On the other hand, in DNA synthesis reaction using DNA polymerase, it is important to set appropriate reaction conditions, as well as to select an appropriate enzyme. Major conditions for reaction include reaction mixture composition, pH, reaction temperature, and template and primer concentration. In addition, these reaction conditions must be set in accordance with the enzyme used and the purpose. However, such settings may be difficult to make in some cases.

In addition, there has been known that efficient DNA synthesis can be carried out by using a combination of plural DNA polymerases, wherein the efficient DNA synthesis could not be achieved by single DNA polymerase [*Proc. Natl. Acad. Sci. USA* 91, 5695-5699 (1994)]. The method is a method using in PCR a mixture of DNA polymerase having 3'→5' exonuclease activity (for example, the above *Pyrococcus furiosus*-derived α-type DNA polymerase) and DNA polymerase not having such an activity (for example, *Thermus aquaticus*-derived DNA polymerase (Taq DNA polymerase), and is known as LA-PCR method. According to this method, there are exhibited such effects that the yield of amplified DNA is increased, as compared with that of conventional PCR using only one kind of DNA polymerase, and that long chain length of DNA which could not be amplified by conventional PCR can be amplified. However, such effects are only exhibited when an enzyme having 3'→5' exonuclease activity is used in combination with an enzyme having no such activity.

As described above, the DNA synthesis reaction using DNA polymerase is indispensable as a genetic engineering procedure. Moreover, it is important to increase their efficiency for research or the like. However, a presently available reaction system has a defect in that it is not sufficiently optimized to be utilized for research or the like. For this reason, there has been a demand for a method enabling more efficient DNA synthesis as compared to that of the conventional DNA synthesis reactions.

#### DISCLOSURE OF INVENTION

The present invention has been accomplished in view of the prior art



described above, and an object of the present invention is to provide (1) a DNA synthesis reaction-enhancer; (2) a DNA synthesis method characterized in that the reaction is carried out in the presence of the DNA synthesis reaction-enhancer; (3) a DNA synthesis reaction composition comprising the DNA synthesis reaction-enhancer; (4) a DNA synthesis reaction composition comprising two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity; (5) a DNA synthesis method characterized in that during the DNA synthesis reaction two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity is used; (6) a kit comprising two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity; and (7) a kit comprising the DNA synthesis reaction-enhancer and DNA polymerase.

As a result of intensive studies, the present inventors have found that efficiency of the DNA synthesis reaction by DNA polymerase is improved in the coexistence of at least one kind selected from the group consisting of acidic substances and cationic complexes. In addition, they have found that extremely efficient DNA synthesis is generated when two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity are mixed. Further, excellent gene amplification reaction system is constructed by combination of these techniques, thereby completing the present invention.

Concretely, the gist of the present invention relates to:

- [1] a DNA synthesis reaction-enhancer comprising at least one kind selected from the group consisting of acidic substances and cationic complexes;
- [2] a DNA synthesis method characterized in that during a DNA synthesis reaction a reaction is carried out in the presence of the DNA synthesis reaction-enhancer of item [1] above by using DNA polymerase;

[3] a DNA synthesis reaction composition comprising the DNA synthesis reaction-enhancer of item [1] above;

[4] a DNA synthesis reaction composition comprising two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity;

5 [5] a DNA synthesis method characterized in that during a DNA synthesis reaction two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity are used;

[6] a kit for use in *in vitro* DNA synthesis, comprising two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity; and

10 [7] a kit for use in *in vitro* DNA synthesis, wherein the kit comprises the DNA synthesis reaction-enhancer of item [1] above and DNA polymerase.

#### BEST MODE FOR CARRYING OUT THE INVENTION

##### (I) DNA Synthesis Reaction-Enhancer of the Present Invention

15 One of the great features of the present invention resides in that the DNA synthesis reaction-enhancer comprises at least one kind selected from the group consisting of acidic substances and cationic complexes. The DNA synthesis reaction-enhancer of the present invention can exhibit an action of enhancing DNA synthesis reaction by DNA polymerase, owing to contain at least one kind  
20 (active ingredient) selected from the group consisting of acidic substances and cationic complexes.

The term "DNA synthesis reaction-enhancer" of the present invention refers to the acidic substance or cationic complex mentioned above alone, or a mixture comprising both the acidic substance and cationic complex, each of  
25 which is capable of exhibiting an action of enhancing DNA synthesis reaction.

In addition, as to the term "DNA synthesis reaction-enhancer" of the present invention, when a complex of an acidic substance and a cationic complex is formed, such a complex is also encompassed therein, as long as the substance is capable of exhibiting an action of enhancing DNA synthesis reaction by DNA polymerase.

Furthermore, mixtures in which various additives are supplemented within the range so that the mixture can exhibit the action of enhancing DNA synthesis reaction by DNA polymerase are also encompassed in the "DNA synthesis reaction-enhancer" of the present invention.

The term "action of enhancing DNA synthesis reaction" can be examined by the chain length of new synthetic DNA chain per unit time or by the amount of amplified product in PCR per unit time. In addition, the action can also be examined by comparing an activity where at least one kind selected from the group consisting of acidic substances and cationic complexes is added with an activity where at least one kind selected from the group consisting of acidic substances and cationic complexes is not added, when determining DNA polymerase activity, for instance, when determining an incorporation of labeled nucleotide into the new synthetic DNA chain. Such an "action of enhancing DNA synthesis reaction" encompasses an action of improving efficiency for synthesis reaction.

It is thought that the action of the DNA synthesis reaction-enhancer of the present invention is in, for example, but are not particularly limited to, that the DNA synthesis reaction-enhancer acts as a DNA polymerase activity-enhancer on DNA polymerase, thereby improving its catalytic activity; or that nonspecific interaction of the enzyme on template DNA is suppressed by retaining DNA

polymerase on a molecule of the active ingredient of the above DNA synthesis reaction-enhancer, to provide the enzyme in the optimum amount for template DNA; and in addition that the DNA synthesis reaction-enhancer acts on template DNA so as to keep its steric structure so that the DNA synthesis reaction is easily progressed, thereby efficiently exhibiting the activity of the DNA polymerase.

In addition, it is thought that another embodiment for the action of the DNA synthesis reaction-enhancer of the present invention is in the improvement of the efficiency of annealing for template DNA and primers.

The acidic substances used in the DNA synthesis reaction-enhancer mentioned above are substances having an action of enhancing DNA synthesis reaction as DNA polymerase activity-enhancers, and concretely include negatively charged substances or salts thereof, especially acidic substances or salts thereof.

In the DNA synthesis reaction-enhancer of the present invention, a DNA synthesis reaction can be enhanced by optimizing the interaction between DNA polymerase and template DNA, which increases with the progress of the DNA synthesis reaction, when an acidic substance is used.

The acidic substances having an action of enhancing DNA synthesis reaction include, for example, but are not particularly limited to, acidic polymer substances such as acidic polysaccharides. In addition, there can be used polyvinyl sulfates, polystyrene sulfates, polyglutamic acids, polyacrylic acids, DNAs not functioning as templates (i.e., DNAs not serving as templates for synthesis of subject DNA), and the like. In the present specification, the term "acidic substance" also encompasses salts thereof. The acidic polysaccharides include, for example, sulfate group-containing sulfated polysaccharides,

representatively exemplified by sulfated-fucose-containing polysaccharides, dextran sulfate, carrageenan, heparin, heparan sulfate, rhamnam sulfate, chondroitin sulfate, dermatan sulfate, and the like, and polyuronic acids such as hyaluronic acid, alginic acid and pectin. In addition, as the sulfated-fucose-containing polysaccharides, there can be used, for example, sulfated-fucose-containing polysaccharide-F or sulfated-fucose-containing polysaccharide-U. The term "sulfated-fucose-containing polysaccharide-F" as used herein refers to a sulfated-fucose-containing polysaccharide substantially contains no uronic acid, obtainable from brown algae or the like by the method, for instance, described in WO 97/26896 or WO 97/47208. In addition, the term "sulfated-fucose-containing polysaccharide-U" refers to a sulfated-fucose-containing polysaccharide containing uronic acid, obtainable by the methods described in the above publications.

The salts of the acidic substances described above are not particularly limited, as long as they have an action of enhancing DNA synthesis reaction, with preference given to water-soluble salts. For example, there are included alkali metal salts such as sodium dextran sulfate, sodium alginate, sodium polyglutamates, sodium polystyrene sulfates, heparin sodium, potassium polyvinyl sulfates, potassium dextran sulfate and heparin lithium.

The acidic substances mentioned above may be those isolated and purified from naturally occurring substances or chemically or enzymatically synthesized products, as long as they maintain an action of enhancing DNA synthesis reaction. In addition, the acidic substances described above may be unpurified or partially purified products containing the acidic substances. Furthermore, the acidic substances described above may be appropriately modified, as long as the

action of enhancing DNA synthesis reaction is maintained. In addition, the acidic substances described above may be substances obtained by degradation procedures so as to have appropriate molecular weights, or those obtained by carrying out molecular weight fractionation after such degradation procedures, as long as these substances have an action of enhancing DNA synthesis reaction. In the present invention, acidic substances having a molecular weight of several thousands or more can be preferably used. Furthermore, these substances may be used singly or in admixture of two or more kinds.

The cationic complexes used for the DNA synthesis reaction-enhancer of the present invention may be any substances, as long as they have an action of enhancing DNA synthesis reaction of DNA polymerase, and concretely include positively charged complexes or salts thereof, especially complex cations or complex salts thereof.

In the DNA synthesis reaction-enhancer of the present invention, when a cationic complex is used, the efficiency for annealing between template DNA and primers can be increased, and as a result, the DNA synthesis reaction can be enhanced. In addition, desired amplified fragments can be obtained even under conditions that have been difficult in DNA amplification, even under conditions of, for example, low magnesium concentrations, low primer concentrations, low enzyme amounts, amplification of GC-rich regions, and the like. Furthermore, when the above cationic complex is used, amplification can be enhanced even in PCR for long-chain DNA.

In addition, the DNA synthesis reaction can be further enhanced by combining the acidic substances and cationic complexes mentioned above.

The cationic complexes having an action of enhancing DNA synthesis

reaction are not particularly limited, and there can be used, for instance, transition metal complexes. In the present specification, the term "cationic complex" also encompasses salts thereof.

The transition metal complex may be any complex capable of exhibiting an action of enhancing DNA synthesis reaction, and complexes having as the central atom a transition element in Group VIII of the Periodic Table (Kagaku Jiten, 1st edition, Tokyo Kagaku Dojin) are preferred. The above Group VIII transition element includes, but not particularly limited, as long as it forms a complex capable of exhibiting an action of enhancing DNA synthesis reaction, for instance, cobalt (Co), rhodium (Rh), iridium (Ir) and the like. In addition, the ligands in the complex include, but not particularly limited to, monodentate ligands, bidentate ligands, tridentate ligands, and tetradentate ligands. For example, neutral ligands such as  $\text{H}_2\text{O}$ ,  $\text{NH}_3$ ,  $\text{CO}$ ,  $\text{NO}$  and pyridine, and chelate ligands such as  $\text{H}_2\text{NCH}_2\text{CH}_2\text{NH}_2$  may be exemplified. In addition, the complex may contain as a ligand an anionic ligand, for instance,  $\text{Cl}^-$ ,  $\text{OH}^-$ ,  $\text{NO}_2^-$ ,  $\text{CN}^-$ ,  $\text{CO}_3^{2-}$ , or the like, as long as it is a cationic complex capable of being present as a complex cation in the reaction mixture. The kinds of ligands in the above complex may be one kind or a plural kinds. Furthermore, as to cationic complexes, there exist geometric isomers, optical isomers and linkage isomers, and any cationic complexes are included in the DNA synthesis reaction-enhancer of the present invention, as long as they have an action of enhancing DNA synthesis reaction.

Concrete examples of the transition metal complex include, for instance,  $[\text{Co}(\text{NH}_3)_6]\text{Cl}_3$ ,  $[\text{Co}(\text{H}_2\text{NCH}_2\text{CH}_2\text{NH}_2)_3]\text{Cl}_3$ ,  $[\text{Co}(\text{NH}_3)_5(\text{H}_2\text{O})]\text{Cl}_3$ ,  $[\text{Co}(\text{H}_2\text{O})(\text{NH}_3)_5]_2(\text{SO}_4)_3$ ,  $[\text{Co}(\text{OH})(\text{NH}_3)_5]\text{Cl}_2$ ,  $[\text{Rh}(\text{H}_2\text{NCH}_2\text{CH}_2\text{NH}_2)_3]\text{Cl}_3$ ,

[Rh(NH<sub>3</sub>)<sub>6</sub>]Br<sub>3</sub>, [Rh(NH<sub>3</sub>)<sub>5</sub>(H<sub>2</sub>O)]Cl<sub>3</sub>, [RhCl<sub>2</sub>(NH<sub>3</sub>)<sub>4</sub>]Cl, [Ir(NH<sub>3</sub>)<sub>6</sub>]Cl<sub>3</sub>,  
 [IrCl(NH<sub>3</sub>)<sub>5</sub>]Cl<sub>2</sub>, [Ir(H<sub>2</sub>NCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>)<sub>3</sub>]I<sub>3</sub>, and the like. Among the above-  
 mentioned transition metal complexes, especially one or more kinds selected  
 from the group consisting of [Co(NH<sub>3</sub>)<sub>6</sub>]Cl<sub>3</sub>, [Co(H<sub>2</sub>NCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>)<sub>3</sub>]Cl<sub>3</sub> and  
 5 [Rh(H<sub>2</sub>NCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>)<sub>3</sub>]Cl<sub>3</sub> are preferable.

The salts of the cationic complexes described above are not particularly  
 limited, as long as they have an action of enhancing DNA synthesis reaction,  
 with preference given to water-soluble salts. For example, there are included  
 salts of halogenides such as chlorides; salts of inorganic acids such as sulfates;  
 10 and salts of organic acids such as acetates. In other words, even salts of the  
 acidic substance and cationic complex of the DNA synthesis reaction-enhancer  
 of the present invention can be preferably used, as long as they ionize in the  
 reaction mixture.

The cationic complex mentioned above may, for example, be any  
 15 substance, as long as it maintains an action of enhancing DNA synthesis reaction.  
 In addition, the cationic complex described above may be an unpurified or  
 partially purified product containing the complex. Furthermore, the cationic  
 complex may be appropriately modified within a range in which an action of  
 enhancing DNA synthesis reaction is maintained. Furthermore, these substances  
 20 can be used singly or in admixture.

The DNA polymerase used for DNA synthesis reaction which is enhanced  
 by the DNA synthesis reaction-enhancer of the present invention is not  
 particularly limited, and includes, for instance, pol I-type DNA polymerases [*E.*  
*coli* DNA polymerase I, Klenow fragment, *Thermus aquaticus*-derived DNA  
 25 polymerase (Taq DNA polymerase), and the like],  $\alpha$ -type DNA polymerases [the



above-mentioned  $\alpha$ -type *Pyrococcus furiosus*-derived DNA polymerase, *Thermococcus litralis*-derived DNA polymerase (VENT DNA polymerase), *Pyrococcus sp.*-derived DNA polymerase (Pyrobest DNA polymerase, KOD DNA polymerase), *Pyrococcus sp.*-derived DNA polymerase GB-D (DEEP VENT DNA polymerase), and the like], and non- $\alpha$ , non-polI type DNA polymerases not belonging to any of these polymerases. Incidentally, each of the polI type DNA polymerases and  $\alpha$ -type DNA polymerases refers to a group of enzymes classified by the homology on the amino acid sequences thereof, and the feature on the amino acid sequences is described in *Nucleic Acids Research* 15, 4045-4057 (1991).

In addition, the non- $\alpha$ , non-pol I type DNA polymerases include, for example, DNA polymerase derived from *Pyrococcus furiosus* described in WO 97/24444. In the present specification, for the sake of distinguishing between the above non- $\alpha$ , non-pol I type DNA polymerase derived from above-mentioned *Pyrococcus furiosus* and the  $\alpha$ -type DNA polymerase, which is similarly produced by *Pyrococcus furiosus*, the non- $\alpha$ , non-pol I type DNA polymerase is referred to as Pfu DNA Polymerase II. In addition, the  $\alpha$ -type DNA polymerase derived from the above-mentioned *Pyrococcus furiosus* is referred to as Pfu DNA Polymerase I.

Pfu DNA Polymerase II is an enzyme having the properties shown below. Properties of Pfu DNA Polymerase II:

- 1) Exhibiting higher activity when polymerase activity is determined for a case of using as the substrate a complex formed by annealing primers to single-stranded template DNA than that of a case of using activated DNA as the substrate;

- 2) Having 3'→5' exonuclease activity;
- 3) Capable of amplifying DNA fragments of about 20 kilo base pairs without addition of another enzyme when polymerase chain reaction (PCR) is carried out with λDNA as a template under the following conditions:

5 Conditions for PCR:

(a) Composition for Reaction Mixture: Comprising 10 mM Tris-hydrochloric acid (pH 9.2), 3.5 mM magnesium chloride, 75 mM potassium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% bovine serum albumin, 0.1% Triton X-100, 5.0 ng/50 µl of λDNA, and 10 pmole/50 µl of primer λA

10 (SEQ ID NO: 1 in Sequence Listing) and primer λB (SEQ ID NO: 2 in Sequence Listing), and 3.7 U/50 µl of DNA polymerase;

(b) Reaction Conditions: PCR is carried out in 30 cycles, wherein one cycle of reaction comprises a process consisting of 98°C, 10 seconds - 68°C, 10 minutes.

15 4) Constituted by two kinds of DNA polymerase-constituting proteins corresponding to about 90,000 Daltons and about 140,000 Daltons on SDS-PAGE.

20 In addition, the gene encoding the above Pfu DNA Polymerase II has been already cloned, and *Escherichia coli* JM109 transformed with plasmid pFU1001 carrying the gene is named and identified as *Escherichia coli* JM109/pFU1001, and deposited under the accession number of FERM BP-5579 with the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, Ministry of International Trade and Industry, of which the

25 address is 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken (Zipcode 305-8566),

Japan, since August 11, 1995 (date of original deposit). Therefore, the transformant is cultured, and Pfu DNA polymerase II mentioned above can be obtained from the resulting culture by, for example, the method described in WO97/24444 (pages 28-29, Example 3).

5 In DNA synthesis reactions using various kinds of DNA polymerases described above, the efficiency of DNA synthesis of the DNA polymerases is surprisingly increased by adding the DNA synthesis reaction-enhancer of the present invention to the reaction mixture. In this case, DNA synthesis efficiency increases similarly when using two or more kinds of DNA polymerases, for  
10 instance, two or more kinds of DNA polymerases each having 3'→5' exonuclease activity (for instance,  $\alpha$ -type DNA polymerase and non- $\alpha$ , non-pol I type DNA polymerase), or one DNA polymerase having 3'→5' exonuclease activity and the other DNA polymerase having no 3'→5' exonuclease activity.

For example, the amount of amplified product increases, when PCR is  
15 carried out by using a reaction mixture supplemented with the DNA synthesis reaction-enhancer of the present invention, in comparison to one obtained by carrying out PCR without this addition. For example, an amplified product can be obtained by the addition of the DNA synthesis reaction-enhancer of the present invention, even under conditions that have conventionally been difficult  
20 in amplification. In addition, the amount of amplified product can be quantified by, for example, subjecting a given amount of the reaction mixture after PCR to electrophoresis, staining the gel after electrophoresis with ethidium bromide or the like, and determining the fluorescence intensity of the band ascribed to the amplified product by means of an imaging analyzer or the like. When the  
25 amount of amplified product is quantified as described above and comparison is

made between one in which PCR is carried out by using a reaction mixture added with the DNA synthesis reaction-enhancer of the present invention and one in which PCR is carried out without such addition, the amount of amplified product shows an increase by about 2 to 5 folds, though the level of increase depends on length of a product to be amplified, GC content, and the like, in comparison with those. In addition, efficient DNA amplification is also possible from a small amount of template DNA.

Furthermore, in PCR using the DNA synthesis reaction-enhancer of the present invention, the reaction time period required for obtaining the same amount of amplified product is shorter as compared to that of conventional PCR.

Generally in PCR, DNA amplification is carried out by three steps of dissociation (denaturation) of double stranded template DNA to single stranded one; annealing of a primer to single stranded template DNA, a complementary strand synthesis (extension) from the primer. In addition, DNA amplification is also carried out in a so-called "shuttle PCR" ["PCR Hou Saizensen (PCR Method Frontier)," *Proteins, Nucleic Acids and Enzymes* an extra number 41, No. 5, 425-428 (1996)], which is a two-step reaction, in which among the three-step reactions described above, the annealing step and the extension reaction step of the primer are carried out at the same temperature. The DNA synthesis reaction-enhancer of the present invention can particularly shorten the time period required for the above extension step, the time period required for an entire synthesis reaction can be shortened, in either of the above-mentioned three-step reaction and two-step reaction.

Here, the reaction time required for obtaining the same amount of amplified product by conventional PCR can be evaluated by, for example, using

as a standard the amount of amplified product when PCR is carried out by using a reaction mixture supplemented with the DNA synthesis reaction-enhancer of the present invention. In other words, the reaction time period can be determined by sampling a reaction mixture for each cycle of PCR, wherein the PCR is

5 carried out without the addition of the above enhancer; quantifying the amount of amplified product as described above; determining the number of cycles until reaching the same amount as the amplified amount obtained in the presence of the DNA synthesis reaction-enhancer of the present invention; and calculating the reaction time period from the number of cycles and the time period needed

10 per one cycle. In addition, the time period needed per one cycle of PCR using the DNA synthesis reaction-enhancer of the present invention can be set to be shorter than the time period needed per one cycle of ordinary PCR. The time period needed per one cycle, in the case of 3-step reactions, for example, can be obtained as the sum of the retention time periods for each step of denaturation, primer annealing, and primer extension, and the time period for reaching each set

15 temperatures from denaturation to primer annealing, from primer annealing to primer extension, and from primer extension to denaturation in the next cycle. In 2-step reactions, the time period needed per one cycle can be obtained as the sum of the retention times for each step and the transition time period between steps.

20 When comparison is made between the total time period needed for the PCR to be carried out using a reaction mixture supplemented with the DNA synthesis reaction-enhancer of the present invention and the total time period needed for PCR to be carried out without the addition, the total time period needed can be shortened to about 1/2, which may vary depending on a length of a fragment to

25 be amplified, GC content, and the like.

Since the DNA synthesis reaction-enhancer of the present invention can shorten the time period required for the entire amplification reaction, there can be exhibited an excellent effect that gene diagnostic method or the like can be performed in shorter time periods by using the enhancer for gene diagnostic method or the like on the basis of PCR method.

## (II) DNA Synthesis Reaction Composition of the Present Invention

As an embodiment, the DNA synthesis reaction composition of the present invention includes, for instance, a composition comprising the DNA synthesis reaction-enhancer described in the above item (I). The above composition may comprise various components necessary for DNA synthesis using DNA polymerase, for example, dNTP, magnesium chloride and buffer components for maintaining appropriate pH. The composition may further comprise DNA polymerase.

The DNA polymerase contained in the DNA synthesis reaction composition is not particularly limited, and includes various kinds of DNA polymerases described in the above item (I). The DNA synthesis reaction composition may contain one kind of enzyme or may contain two or more kinds of (plural kinds of) enzymes. For example, when a plurality of DNA polymerases are contained, the DNA polymerase may be the combination of DNA polymerase having 3'→5' exonuclease activity used for LA-PCR and DNA polymerase having no such activity mentioned above. In addition, the DNA synthesis reaction composition of the present invention comprising thermostable DNA polymerase is suitable for synthesis of DNAs having nucleotide sequences which easily form higher order structures and the use for

PCR.

In addition, another embodiment of the DNA synthesis reaction composition of the present invention includes a composition comprising two or more kinds of DNA polymerases each having 3'→5' exonuclease activity. The above composition may also comprise various components necessary for DNA synthesis as described above and/or the DNA synthesis reaction-enhancer mentioned above.

As a composition comprising a plurality of DNA polymerases, compositions utilized in LA-PCR method have been conventionally known. As described above, a composition utilized for LA-PCR method is prepared by combining a DNA polymerase having 3'→5' exonuclease activity and a DNA polymerase having no such activity or exhibiting no such activity.

The term "having 3'→5' exonuclease activity" as used herein means that the 3'→5' exonuclease activity owned by naturally occurring DNA polymerase has not been removed or reduced by a chemical method or genetic engineering method, i.e., the activity is substantially owned. In addition, the term "DNA polymerase exhibiting no 3'→5' exonuclease activity" refers to a DNA polymerase of which 3'→5' exonuclease activity has been removed or reduced by a chemical method or genetic engineering method.

The present inventors have clarified that when DNA polymerases each having 3'→5' exonuclease activity are combined, surprisingly, DNA synthesis reaction can be accomplished at higher reaction rates than those by LA-PCR method.

Such compositions include, for instance, but are not particularly limited to, a composition comprising a combination of thermostable DNA polymerase

belonging to the  $\alpha$ -type and thermostable DNA polymerase belonging to the non- $\alpha$ , non-pol I type. In addition, the performance of the above composition is improved by the addition of the DNA synthesis reaction-enhancer described above.

5           One of the features of the above composition resides in that the DNA synthesizing rate per unit time is very high. When the above composition is used for PCR method, the time period needed for amplifying DNA of the same chain length is shorter than that for conventional PCR method and LA-PCR method. Therefore, DNA amplification can be carried out even under PCR conditions that  
10           were impossible for DNA amplification by conventional methods.

          Since the DNA synthesis reaction composition of the present invention can shorten the time period needed for an entire amplification reaction, there is exhibited an excellent effect that gene diagnostic method or the like can be performed in shorter time periods by using the composition, for instance, for  
15           genetic diagnostic method or the like on the basis of PCR method.

### (III) DNA Synthesis Method of the Present Invention

          According to the use of the DNA synthesis reaction-enhancer of the present invention or the DNA synthesis reaction composition of the present  
20           invention, a DNA synthesis reaction can be carried out at higher efficiency than that of conventional method by preparing a reaction mixture. For example, when the DNA synthesis method of the present invention is used for PCR method, an amplified product can be obtained with shorter reaction time than conventional PCR method or LA-PCR method.

25           Embodiments for the DNA synthesis method of the present invention



include a method using two or more kinds of DNA polymerases; a method using DNA polymerase having 3'→5' exonuclease activity and DNA polymerase having no such activity; a method using two or more kinds of DNA polymerases each having 3'→5' exonuclease activity; and a method using α-type DNA polymerase and non-α, non-pol I type DNA polymerase. Further, there is included a method in which the DNA synthesis method mentioned above is carried out by PCR method.

According to the DNA synthesis method of the present invention, since the time period needed for an entire amplification reaction can be shortened, there can be exhibited an excellent effect that gene diagnostic method or the like can be performed in shorter time periods by using the method for gene diagnostic method or the like on the basis of PCR.

In addition, the DNA synthesis method of the present invention can also be utilized for such procedures utilizing DNA polymerases as DNA labeling and nucleotide sequencing by the dideoxy method.

#### (IV) Kits for DNA Synthesis Method of the Present Invention

DNA can be synthesized more conveniently and efficiently by using the kit of the present invention. The kit is not particularly limited, as long as the kit is usable for reactions involving DNA synthesis, and includes kits for DNA synthesis reactions *in vitro*. Concrete examples include kits for DNA nucleotide sequencing by the dideoxy method, kits for DNA labeling, kits for PCR, kits for cDNA synthesis, and kits for site-directed mutagenesis.

One of the great features of the kit of the present invention resides in that the kit comprises the DNA synthesis reaction-enhancer of the present invention

and/or two or more kinds of DNA polymerases each having 3'→5' exonuclease activity. In other words, an embodiment includes a kit comprising the DNA synthesis reaction-enhancer of the present invention, and another embodiment includes a kit comprising two or more kinds of DNA polymerases each having 3'→5' exonuclease activity. Furthermore, the present invention also encompasses a kit comprising both the DNA synthesis reaction-enhancer of the present invention and the two or more kinds of DNA polymerases each having 3'→5' exonuclease activity.

The DNA synthesis reaction enhancers include at least one kind selected from the group consisting of acidic substances and cationic complexes described in the above item (I). The two or more kinds of DNA polymerases each having 3'→5' exonuclease activity include, for instance,  $\alpha$ -type DNA polymerase and non- $\alpha$ , non-pol I type DNA polymerase, the polymerases preferably being thermostable DNA polymerases. The kit comprising the DNA synthesis reaction-enhancer of the present invention further comprises one kind or two or more kinds of appropriate DNA polymerases, with particular preference given to thermostable DNA polymerases. This kit may also comprise reagents necessary for DNA polymerase reactions, such as dNTP, magnesium chloride, and buffer components for keeping the reaction mixture at appropriate pH, in any of the embodiments.

The DNA synthesis reaction-enhancer mentioned above and DNA polymerases may be contained in the kit in the form of single components, or in the form added to the reaction buffer or the like.

Since the DNA synthesis reaction using the kit of the present invention allows to give a very large amount of DNA synthesized per unit time period than

that of ordinary reaction systems, there is exhibited an excellent effect that the time period needed for the procedures can be shortened when applied to various kinds of procedures involving DNA synthesis reactions, representatively exemplified by DNA labeling, PCR, cDNA synthesis, and site-directed mutagenesis. For example, when this kit is applied to PCR, the time period needed for amplifying a DNA of the same chain length is shorter than that of conventional PCR method and LA-PCR method. Therefore, DNA amplification can be carried out even under PCR conditions that were impossible for DNA amplification by conventional methods. In addition, the kit of the present invention can shorten the time period needed for an entire amplification reaction, there is exhibited an excellent effect that gene diagnostic method or the like can be performed in shorter time periods by using the kit for gene diagnostic method or the like on the basis of PCR method.

In addition, the kit of the present invention is utilized to amplify nucleic acids by PCR method, so that there is exhibited an excellent effect that the time period needed for an entire amplification reaction can be shortened, or that the yield of the product obtained by the amplification reaction can be improved. In this case, a reaction mixture for amplification is prepared by mixing reagents necessary for PCR contained in the kit, and at least one kind selected from the group consisting of acidic substances and cationic complexes is added to the reaction mixture. Thereafter, a reaction in cycles is carried out in which each of denaturation, annealing, and extension processes is repeated. At least one kind selected from the group consisting of acidic substances and cationic complexes is added in an amount effective for shortening the reaction time period and/or for increasing the yield of the amplified product. Furthermore, in PCR method

utilizing the kit of the present invention, there is exhibited an excellent effect that the chain length of the fragment amplified is made longer, in comparison to that obtained by PCR method not utilizing the kit of the present invention.

The reaction mixture mentioned above comprises the following components: 1) a nucleic acid serving as a template, 2) an appropriate reaction buffer, 3) DNA polymerase, 4) dATP, dTTP, dGTP and dCTP, and 5) at least one oligonucleotide primer. At least one kind selected from the group consisting of acidic substances and cationic complexes may be added in an effective amount to the reaction mixture after preparation, or may be added as a component of the reaction buffer.

According to the present invention, there is exhibited an excellent effect gene diagnostic method or the like can be performed in shorter time periods by the use for gene diagnostic method or the like on the basis of PCR, which involves the handling of a large number of samples.

The present invention will be hereinbelow described in further detail by means of the working examples, without intending to restrict the scope of the present invention to these working examples.

In the following working examples, the activities of the commercially available enzymes were shown on the basis of the indication for each enzyme, provided that the enzyme activity unit for Pfu DNA Polymerase II was shown by the value obtained by the method for determination of an enzyme activity described in Example 1. In addition, unless specified otherwise, the reaction solution comprising a commercially available enzyme was prepared in accordance with the instruction for each enzyme, or prepared by using a reaction

buffer attached thereto. Unless specified otherwise, PCR was carried out by using GeneAmp PCR System 9600 (manufactured by Perkin-Elmer).

#### Example 1: Preparation of Pfu DNA Polymerase II

5 Pfu DNA Polymerase II was purified from bacterial cells obtained by culturing *Escherichia coli* JM109/pFU1001 (FERM BP-5579), which was *Escherichia coli* JM109 transformed with plasmid pFU1001 carrying a gene encoding Pfu DNA Polymerase II, and used in the following procedures. Here, the culture of the transformant and the purification of the enzyme were carried  
10 out in accordance with the method described in WO97/24444 (for instance, pages 28-29, Example 3, and the like).

Incidentally, the enzyme activity of Pfu DNA Polymerase II was determined in the following manner. An activated calf thymus DNA (manufactured by Worthington) (activated DNA) was used as a substrate. DNA  
15 activation and determination of DNA polymerase activity were carried out by the method described in *DNA Polymerase from Escherichia coli*, 263-276 (authored by C.C. Richardson), published by Harper & Row, edited by D. R. Davis. To 5 µl of a sample of which the activity was to be determined was added 45 µl of a reaction mixture [20 mM Tris-hydrochloric acid (pH 9.0), 15 mM magnesium  
20 chloride, 2 mM 2-mercaptoethanol, 0.2 mg/ml activated DNA, 40 µM each of dATP, dCTP, dGTP and dTTP, 60 nM [<sup>3</sup>H]-dTTP (manufactured by Amersham)]. The resulting mixture was reacted at 75°C for 5 minutes. A 40 µl portion of this reaction mixture was then spotted onto DE paper (manufactured by Whatman) and washed with 5% by weight Na<sub>2</sub>HPO<sub>4</sub> five times. Thereafter,  
25 the remaining radioactivity on the DE paper was determined using a liquid

scintillation counter. The amount of enzyme which incorporated 10 nmol of [<sup>3</sup>H]-dTTP per 30 minutes into the substrate DNA, determined by the above-described enzyme activity determination method, was defined as 1 U of the enzyme.

5

#### Example 2: Preparation of Primers

Eleven kinds of primers  $\lambda$ 1 to  $\lambda$ 5,  $\lambda$ 7 to  $\lambda$ 10 and  $\lambda$ L36 and  $\lambda$ R485 were synthesized on the basis of the nucleotide sequences of  $\lambda$ DNA. The nucleotide sequences for the primers  $\lambda$ 1 to  $\lambda$ 5 and  $\lambda$ 8 to  $\lambda$ 10 are respectively shown in SEQ ID NOs: 3 to 10 of Sequence Listing. In addition, the nucleotide sequence for  $\lambda$ 7 is shown in SEQ ID NO: 11 of Sequence Listing. Further, the nucleotide sequences for  $\lambda$ L36 and  $\lambda$ R485 are respectively shown in SEQ ID NOs: 12 to 13 of Sequence Listing. The chain lengths of amplified DNA fragments are shown in Table 1, the fragments being amplified by PCR with  $\lambda$ DNA as a template using the combinations of these primers. Further, 3 kinds of primers Eco1, Eco2 and Eco5 were synthesized on the basis of the nucleotide sequence of *Escherichia coli* genomic DNA. The nucleotide sequences of Eco1, Eco2 and Eco5 are respectively shown in SEQ ID NOs: 14 to 16 of Sequence Listing. The chain lengths of amplified DNA fragments are shown in Table 1, the fragments being amplified by PCR with *E. coli* genomic DNA as a template using the combinations of these primers. Further, 2 kinds of primers ApoE-1 and ApoE-2 were synthesized on the basis of the nucleotide sequence of human ApoE region. The nucleotide sequences of ApoE-1 and ApoE-2 are respectively shown in SEQ ID NOs: 17 to 18 of Sequence Listing. The chain lengths of amplified DNA fragments are shown in Table 1, the fragments being amplified by PCR

with human genomic DNA as a template using the combinations of these primers.

Table 1

Primer Pair	Chain Length of Amplified DNA Fragment (kb)
$\lambda 1/\lambda 2$	0.5
$\lambda 1/\lambda 3$	1
$\lambda 1/\lambda 4$	2
$\lambda 1/\lambda 5$	4
$\lambda 1/\lambda 7$	8
$\lambda 1/\lambda 8$	10
$\lambda 1/\lambda 9$	12
$\lambda 1/\lambda 10$	15
$\lambda L36/\lambda R485$	48.5
Eco1/Eco2	2
Eco1/Eco5	10
ApoE- 1/ApoE-2	0.44

### Example 3: Enhancement of DNA Synthesis Reaction by Acidic Substance

#### 5 (1) Effects on Activity of Pfu DNA Polymerase II

The sulfated-fucose-containing polysaccharide-F obtained by the method described in WO97/26896 (for instance, Example 7, page 77, line 8 to page 78, line 13; Example 9, page 79, lines 7 to 18, and the like), the calf thymus DNA (manufactured by Worthington), heparin (manufactured by Wako Pure  
10 Chemicals) were used as acidic substances to examine the effects of these acidic substances imparted on the activity of Pfu DNA Polymerase II.

A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda 1$  and  $\lambda 3$  as a primer pair, and Pfu DNA Polymerase II as DNA polymerase was prepared,

and PCR was carried out. The composition of the reaction mixture is as follows.

#### Composition of Reaction Mixture:

10 mM Tris-hydrochloric acid (pH 9.2), 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01%  
 5 BSA, 1.25 U of Pfu DNA Polymerase II, 500 pg of  $\lambda$ DNA, and 5 pmol each of primers  $\lambda$ 1 and  $\lambda$ 3 (a final volume being 25  $\mu$ l).

Further, 5 ng of the sulfated-fucose-containing polysaccharide-F, 200 ng of the calf thymus DNA, or 0.5 ng, 1 ng, 2 ng or 5 ng of heparin was respectively added to the above reaction mixture.

10 The reaction was carried out in 25 cycles, wherein one cycle of reaction comprises a process consisting of 98°C, 0 seconds - 68°C, 0 seconds. Here, in the present specification, the description such as "98°C, 0 seconds," "68°C, 0 seconds" shows that the reactor is programmed so that the shift to the next temperature setting takes place simultaneously as the temperature reaches the set  
 15 temperature.

After the termination of reaction, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%. The gel after electrophoresis was subjected to ultraviolet irradiation, to visualize a band ascribed to the amplified fragment,  
 20 whereby an amplified fragment was confirmed. As a result, there was confirmed that an expected fragment of 1 kb was excellently amplified in any case where the acidic substances were added. When 0.5 ng of heparin was added, the fragment was especially highly efficiently amplified. On the other hand, for a similar reaction mixture except that an acidic substance was not added, when  
 25 PCR was carried out under the above conditions, an amplified fragment of 1 kb



could not be confirmed.

Further, the studies on the amplification of long chain length of DNA were carried out. A similar reaction mixture for PCR as above except that the amount of  $\lambda$ DNA as a template was changed to 2.5 ng, and that the primer pair was changed to primers  $\lambda$ 1 and  $\lambda$ 5 or primers  $\lambda$ 1 and  $\lambda$ 10, respectively, was prepared. Here, 200 ng of the calf thymus DNA or 5 ng of the sulfated-fucose-containing polysaccharide-F was added as an acidic substance. The reaction was carried out in 30 cycles, wherein one cycle of reaction comprises a process consisting of 98°C, 0 seconds - 68°C, 5 minutes. As a result, when the calf thymus DNA or the sulfated-fucose-containing polysaccharide-F was added, an amplified fragment of 4 kb was confirmed for the primers  $\lambda$ 1 and  $\lambda$ 5, and an amplified fragment of 15 kb was confirmed for primers  $\lambda$ 1 and  $\lambda$ 10, respectively. On the other hand, in a case where these acidic substances were not added, amplified fragments could not be confirmed with either of the primer pairs.

## (2) Effects of Pfu DNA Polymerase I on DNA Synthesis Reaction

The effects of the sulfated-fucose-containing polysaccharide-F, which is an acidic substance, imparted on the activity of Pfu DNA Polymerase I were examined. Here, an enzyme prepared by utilizing recombinant DNA technique (Cloned Pfu DNA Polymerase, manufactured by Stratagene) was used for Pfu DNA Polymerase I.

A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda$ 1 and  $\lambda$ 2 as a primer pair, and Pfu DNA Polymerase I as DNA polymerase was prepared, and PCR was carried out. The composition of the reaction mixture is as follows.

Composition of Reaction Mixture:

Pfu DNA Polymerase I buffer (manufactured by Stratagene), 0.2 mM each of dATP, dCTP, dGTP and dTTP, 1.25 U of Pfu DNA Polymerase I, 500 pg of template DNA, and 5 pmol each of primers  $\lambda 1$  and  $\lambda 2$  (a final volume being 25  $\mu$ l). Two kinds of reaction mixtures were prepared: One with the addition of 20 ng of the sulfated-fucose-containing polysaccharide-F to the reaction mixture, and one without addition.

The reaction in 25 cycles was carried out for the prepared reaction mixture, wherein one cycle of reaction comprises a process consisting of 94°C, 30 seconds - 55°C, 30 seconds - 72°C, 60 seconds. Thereafter, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel, whereby an amplified DNA fragment was confirmed. The amount of the amplified DNA fragment of 0.5 kb was visualized in the same manner as described above, and compared. As a result, it was found that the amount of DNA amplified in the reaction mixture added with sulfated-fucose-containing polysaccharide-F was larger. From this finding, it was confirmed that the DNA amplification efficiency by Pfu DNA Polymerase I is improved by the addition of the sulfated-fucose-containing polysaccharide-F.

#### Example 4: PCR Using Two Kinds of DNA Polymerases Each Having 3' $\rightarrow$ 5'

##### Exonuclease Activity

Studies were made on PCR simultaneously using both Pfu DNA Polymerase II and Pfu DNA Polymerase I, each of which is DNA polymerase having 3'  $\rightarrow$  5' exonuclease activity. Here, Cloned Pfu DNA Polymerase (manufactured by Stratagene) mentioned above was used for Pfu DNA Polymerase I.

# (1) PCR Using Mixture of Pfu DNA Polymerase I and Pfu DNA Polymerase II

A reaction mixture comprising  $\lambda$ DNA as a template, and primers  $\lambda$ 1 and  $\lambda$ 8 as a primer pair, was prepared, and PCR was carried out. In this PCR, both Pfu DNA Polymerase I and Pfu DNA Polymerase II were used. The composition of the reaction mixture is as follows.

## Composition of Reaction Mixture:

10 mM Tris-hydrochloric acid (pH 9.2), 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 500 pg of template DNA, and 5 pmol each of primers  $\lambda$ 1 and  $\lambda$ 8

To the reaction mixture were added 0.375 U of Pfu DNA Polymerase II, and 0.125, 0.375, 0.75 or 1.25 U of Pfu DNA Polymerase I, respectively, to make up a final volume of the reaction mixture of 25  $\mu$ l. In addition, as controls, a reaction mixture prepared by adding only Pfu DNA Polymerase I, and a reaction mixture prepared by adding only Pfu DNA Polymerase II were also prepared.

The reaction in 25 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 0 seconds - 68°C, 3 minutes. Thereafter, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. As a result, amplification of a DNA fragment of 10 kb could not be confirmed by the addition of Pfu DNA Polymerase I alone regardless of the amount thereof. In addition, while a very slight amount of a fragment of 10 kb could be found by

the addition of Pfu DNA Polymerase II alone, it was confirmed that amplification efficiency of a fragment of 10 kb was improved in an amount-dependent manner by adding Pfu DNA Polymerase I to Pfu DNA Polymerase II.

## 5 (2) Amplification of DNA Fragments Having Different Chain Lengths

Using  $\lambda$ DNA as a template, and each of primers  $\lambda$ 1 and  $\lambda$ 3, primers  $\lambda$ 1 and  $\lambda$ 4, or primers  $\lambda$ 1 and  $\lambda$ 5 as a primer pair, the reaction mixture having the following composition was prepared.

### Composition of Reaction Mixture:

10 10 mM Tris-hydrochloric acid (pH 9.2), 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 1.25 U of Pfu DNA Polymerase I and 1.25 U of Pfu DNA Polymerase II, 500 pg of  $\lambda$ DNA, and 5 pmol each of the primers, 5 ng or 50 ng of sulfated-fucose-containing polysaccharide-F (a final volume being 25  $\mu$ l). In addition, as  
15 a control, one without addition of sulfated-fucose-containing polysaccharide-F was prepared.

The reaction in 30 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 0 seconds - 68°C, 0 seconds. Thereafter, 5  $\mu$ l of the resulting reaction mixture  
20 was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. As a result, amplification of a DNA fragment of 1 kb, 2 kb or 4 kb could be confirmed, depending upon the primer pair used. Regarding sulfated-fucose-containing polysaccharide-F, one added in an amount of 50 ng showed more excellent  
25 amplification efficiency as compared to the one added in an amount of 5 ng. On

the other hand, in a case of one without addition, amplified fragments could not be confirmed.

#### Example 5: Comparison with Conventional PCR Techniques

##### 5 (1) Amplification of Long Chain DNA

As to the amplification reaction of a DNA fragment of 10 to 15 kb using  $\lambda$ DNA as a template, a comparison was made between the reaction system described in item (2) of Example 4 and that for LA-PCR method using a combination of DNA polymerase having 3'  $\rightarrow$  5' exonuclease activity and DNA  
10 polymerase having no such activity. Here, the reaction mixture for LA-PCR was prepared by using TaKaRa LA PCR Kit Ver. 2 (manufactured by Takara Shuzo Co., Ltd.).

Using each of primers  $\lambda$ 1 and  $\lambda$ 8, primers  $\lambda$ 1 and  $\lambda$ 9, or primers  $\lambda$ 1 and  $\lambda$ 10 as a primer pair, the reaction mixture having the following composition was  
15 prepared.

##### Composition of Reaction Mixture:

10 mM Tris-hydrochloric acid (pH 9.2), 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 1.25 U of Pfu DNA Polymerase I and 1.25 U of Pfu DNA Polymerase II,  
20 500 pg of  $\lambda$ DNA, and 5 pmol each of the primers, 50 ng of sulfated-fucose-containing polysaccharide-F (a final volume being 25  $\mu$ l).

In addition, a reaction mixture for LA-PCR (a final volume being 25  $\mu$ l) has the same amounts of template DNA and primers as the above-mentioned reaction mixture was prepared by using TaKaRa LA PCR Kit Ver. 2 mentioned  
25 above.

The reaction in 25 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 0 seconds - 68°C, 3 minutes. Thereafter, 5 µl of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. As a result, when Pfu DNA Polymerase I and Pfu DNA Polymerase II described in item (2) of Example 5 were used in combination, amplification of a DNA fragment of each of 10 kb, 12 kb and 15 kb for each primer pair was confirmed.

On the other hand, in the reaction employing the conventional LA-PCR, while amplification of a fragment of 10 kb was confirmed, amplification of fragments of 12 kb and 15 kb could not be found.

Incidentally, when amplification of the fragment of 12 kb was tried by changing the conditions for one cycle to one comprising a process consisting of 98°C, 0 seconds - 68°C, 5 minutes, amplification was found also for the reaction mixture for LA-PCR in this case.

## (2) Amplification by Short Time Reaction of Short Chain DNA

As to the amplification reaction of a DNA fragment of 0.5 to 4 kb using λDNA as a template, a comparison was made between the reaction system described in item (2) of Example 4 and that for LA-PCR method or that for KOD DNA Polymerase, which has been known to have high DNA synthesizing rate. Here, the reaction mixture for LA-PCR was prepared by using TaKaRa LA PCR Kit Ver. 2, and the reaction mixture for KOD DNA Polymerase was prepared by using KOD DNA Polymerase, manufactured by TOYOBO CO., LTD. and attached reaction buffer, respectively.

Using each of primers  $\lambda 1$  and  $\lambda 2$ , primers  $\lambda 1$  and  $\lambda 3$ , or primers  $\lambda 1$  and  $\lambda 5$  as a primer pair, the reaction mixture having the following composition was prepared.

Composition of Reaction Mixture:

5 10 mM Tris-hydrochloric acid (pH 9.2), 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 1.25 U of Pfu DNA Polymerase I and 1.25 U of Pfu DNA Polymerase II, 500 pg of  $\lambda$ DNA, and 5 pmol each of the primers, 50 ng of sulfated-fucose-containing polysaccharide-F (a final volume being 25  $\mu$ l).

10 In addition, using the same primer pair as the above-mentioned reaction mixture, a reaction mixture for LA-PCR and a reaction mixture for KOD DNA Polymerase (a final volume being both 25  $\mu$ l) were prepared. Here, to these two kinds of the reaction mixtures, template  $\lambda$ DNA was added 2500 pg each.

The reaction in 25 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 0 seconds - 68°C, 0 seconds. Thereafter, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. As a result, when Pfu DNA Polymerase I and Pfu DNA Polymerase II described in item (2) of Example 5 were used in combination, amplification of a DNA fragment of each of 0.5 kb, 1 kb and 4 kb for each primer pair was confirmed.

25 On the other hand, in the conventional reaction mixture for LA-PCR or reaction mixture for KOD DNA Polymerase, only slight amplification of a DNA fragment of 0.5 kb could be found, even though the reaction mixtures contained template DNA in a five-fold amount.

### (3) Comparison of DNA Detection Sensitivity

As to the amplification reaction of a DNA fragment using a slight amount of  $\lambda$ DNA as a template, a comparison was made between the reaction system described in item (2) of Example 4 and that for LA-PCR method. Here, the reaction mixture for LA-PCR was prepared by using TaKaRa LA PCR Kit Ver. 2.

Using primers  $\lambda 1$  and  $\lambda 2$  as a primer pair, the reaction mixture having the following composition was prepared.

#### Composition of Reaction Mixture:

10 mM Tris-hydrochloric acid (pH 9.2), 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 1.25 U of Pfu DNA Polymerase I and 1.25 U of Pfu DNA Polymerase II, 0.05 fg of  $\lambda$ DNA, and 5 pmol each of the primers, and 50 ng of sulfated-fucose-containing polysaccharide-F (a final volume being 25  $\mu$ l).

In addition, a reaction mixture for LA-PCR having the same amounts of template DNA and primers as the above-mentioned reaction mixture was prepared.

The reaction in 50 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 0 seconds - 68°C, 0 seconds. Thereafter, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. As a result, amplification of a DNA fragment of 0.5 kb could be confirmed only in the reaction system described in item (2) of Example 4.

Next, the primer pairs were changed to primers  $\lambda 1$  and  $\lambda 8$ , and the two



kinds of reaction mixtures in the same manner as mentioned above were similarly prepared. The reaction in 50 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 0 seconds - 68°C, 3 minutes. Thereafter, 5 µl of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. As a result, amplification of a DNA fragment of 10 kb could be found in the reaction system described in item (2) of Example 4, whereas no amplification of a DNA fragment was found in the conventional reaction mixture for LA-PCR.

Example 6: Preparation of Sulfated-fucose-containing Polysaccharide-F by Extraction with Hot Water

The sulfated-fucose-containing polysaccharide-F used in the subsequent examples was purified by the following process. A preparation example of sulfated-fucose-containing polysaccharide-F is given hereinbelow.

Gagome kombu was sufficiently dried, and thereafter 20 kg, a weight on a dry basis, of the Gagome kombu was pulverized. Next, the resulting dry powder was suspended in 900 liters of tap water containing 7.3 kg of calcium chloride•dihydrate, and the temperature was raised to 90°C over a period of 40 minutes with stirring. The extraction was carried out for one hour with keeping at 90° to 95°C. Thereafter, the resulting solution was cooled to 20°C, stopped stirring, and allowed to stand overnight, to give an extract.

Next, solid-liquid separation was carried out using a centrifuge (Model CNA, manufactured by Westfalia Separator Inc.). About 900 liters of

supernatant of the solid-liquid separation was obtained from the above extract by using the centrifuge. Three-hundred and sixty liters of the supernatant was filtered with a SPARKLER FILTER (manufactured by Nippon Senshoku Kikai) incorporated with a filter of a size of 3  $\mu$ m (manufactured by Nippon Shokuhin Rozai). The filtrate was concentrated to a volume of 20 liters by UF membrane ("FE10-FC-FUS0382," manufactured by DAICEL CHEMICAL INDUSTRIES, LTD.) having a fractionated molecular weight of 30000. Thereafter, 20 liters of tap water was added to the resulting concentrate, and the dilution was concentrated again to a volume of 20 liters. The dilution-concentration operations as described above were repeated five time, to give 25 liters of a concentrate.

To 700 ml of the above concentrate were added 0.2 M calcium chloride and 20 mM sodium acetate as a final concentration. Thereafter, the resulting mixture was dialyzed against 20 mM sodium acetate-equilibrated buffer (pH 6.0) containing 0.2 M calcium chloride. The solution after the dialysis treatment was applied to 3500 ml of DEAE-Sepharose FF column (column inner diameter: 9.7 cm) equilibrated with 10 liters of the above equilibrated buffer, and washed with 5 liters of the equilibrated buffer. The elution was carried out under 3-step gradient conditions given hereinbelow.

Incidentally, the flow rate of the chromatography was set at 3500 ml/1 hour. Gradient Conditions:

- 1] linear gradient of 0 to 0.5 M sodium chloride  
(amount of eluent: 4.5 liters)
- 2] linear gradient of 0.5 to 1.0 M sodium chloride  
(amount of eluent: 4.5 liters)

- 3] linear gradient of 1.0 to 2.0 M sodium chloride  
(amount of eluent: 4.5 liters)

5 The eluent was collected 250 ml per one fraction. Each fraction was  
subjected to sugar quantification by phenol sulfuric acid method, and to uronic  
acid quantification by carbazole-sulfuric acid method. As a result, fractions of  
Fraction Nos. 40 to 53, which were fractions having high sugar contents and low  
contents of uronic acid, were obtained. The fractions of Fraction Nos. 40 to 53  
are referred to "sulfated-fucose-containing polysaccharide-F fractions." Each of  
10 the sulfated-fucose-containing polysaccharide-F fractions was concentrated with  
a ultrafiltration membrane of 100000, and thereafter the concentrate was  
dialyzed against 50 mM sodium citrate, and further dialyzed overnight against  
distilled water. Subsequently, the mixture was lyophilized, to give 1.696 g of  
sulfated-fucose-containing polysaccharide-F from the sulfated-fucose-containing  
15 polysaccharide-F fraction.

Example 7: Preparation of Sulfated-fucose-containing Polysaccharide-U by  
Extraction with Hot Water

20 The sulfated-fucose-containing polysaccharide-U used in the subsequent  
examples was purified by the following process. A preparation example of  
sulfated-fucose-containing polysaccharide-U is given hereinbelow.

Two tons of Gagome seaweed was pulverized, and the resulting dry  
powder was suspended in 44 kl of tap water containing 730 kg of calcium  
chloride•dihydrate, and the temperature was raised to 95°C over a period of  
25 1 hour and 35 minutes with stirring. The extraction was carried out for 2 hours

with keeping at 95°C. Thereafter, the resulting solution was cooled overnight to room temperature.

Next, solid-liquid separation was carried out by using a decanter (manufactured by Tanabe Wiltech). The operating conditions were such that the separation was carried out at 3000 to 3300 rpm (2560 G) over a period of about 8 hours. The resulting supernatant of the solid-liquid separation was concentrated to a volume of 5 liters with a UF membrane ("FE10-FC-FUS0382," manufactured by DAICEL CHEMICAL INDUSTRIES, LTD.) having a fractionated molecular weight of 30000. Thereafter, the concentrate was subjected to UF desalting.

The above desalted solution was subjected to turbid removal with LINT FILTER (manufactured by Naigai Joki) incorporated with ADVANTEC #327 filter paper (manufactured by ADVANTEC). The filtrate was sterilized at 98°C for 40 seconds with a plate heater (manufactured by Hisaka Seisakusho). The resulting extract was about 4620 liters. A part of the extract was lyophilized with a lyophilizer (manufactured by Kyowa Shinku Gijutsu).

Three-hundred and forty grams of the lyophilized product was dissolved in 21.2 liters of 20 mM sodium acetate-equilibrated buffer (pH 6.0) containing 0.2 M calcium chloride. The solution was applied to about 35 liters of DEAE-Sepharose FF column (column inner diameter: 35 cm) equilibrated with the above equilibrated buffer, and washed with 70 liters of the equilibrated buffer. The elution was carried out under 3-step gradient conditions given hereinbelow.

Incidentally, the flow rate of the chromatography was set at 35 liters/1 hour. Gradient Conditions:

1] linear gradient of 0 to 0.5 M sodium chloride

(amount of eluent: 120 liters)

2] linear gradient of 0.5 to 1.0 M sodium chloride

(amount of eluent: 120 liters)

3] linear gradient of 1.0 to 1.5 M sodium chloride

5 (amount of eluent: 120 liters)

10 The eluent was collected 10 liters per one fraction. Each fraction was subjected to sugar quantification by phenol sulfuric acid method, and to uronic acid quantification by carbazole-sulfuric acid method. As a result, fractions of Fraction Nos. 4 to 16, which were fractions having high sugar contents and high contents of uronic acid were obtained. The fractions of Fraction Nos. 4 to 16 are referred to "sulfated-fucose-containing polysaccharide-U fractions." Each of the sulfated-fucose-containing polysaccharide-U fractions was concentrated with a ultrafiltration membrane having a cut-off molecular weight of 10000. Thereafter, 15 the resulting concentrate was dialyzed by using a cellulose tube having a cut-off molecular weight of 12000 to 14000. First, the dialysis was carried out by twice an operation of exchanging an aqueous solution of 50 mM trisodium citrate•dihydrate every 3 to 4 hours with the aqueous solution, and thereafter overnight against the aqueous solution. Next, the dialysis was carried out by 20 twice an operation of exchanging distilled water every 3 to 4 hours with distilled water, and thereafter overnight against distilled water. Subsequently, the mixture was lyophilized, to give 37.6 g of sulfated-fucose-containing polysaccharide-U from the sulfated-fucose-containing polysaccharide-U fraction.

25 Example 8: Enhancement of DNA Synthesis Reaction of Pfu DNA Polymerase

## II by Acidic Substances

In the following example, PCR was carried out in Fast mode by using TaKaRa PCR Thermal Cycler Personal (manufactured by Takara Shuzo Co., Ltd.).

### 5 (1) Effects of Sulfated-fucose-containing Polysaccharide-F, Dextran Sulfate Powder and Sodium Alginate on Activity of Pfu DNA Polymerase II

Using the sulfated-fucose-containing Polysaccharide-F obtained in Example 6 described above, dextran sulfate powder (manufactured by Onco), or sodium alginate (100 to 150 centipoises, manufactured by Wako Pure  
10 Chemicals), the effects of these acidic substances on the activity of Pfu DNA Polymerase II were examined. A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda$ 1 and  $\lambda$ 2 as a primer pair, and Pfu DNA Polymerase II as DNA polymerase was prepared, and PCR was carried out. The composition of the reaction mixture is as follows.

#### 15 Composition of Reaction Mixture:

10 mM Tris-hydrochloric acid, pH 9.2, 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 1.25 U of Pfu DNA Polymerase II, 100 pg of  $\lambda$ DNA, and 5 pmol each of primers  $\lambda$ 1 and  $\lambda$ 2 (a final volume being 25  $\mu$ l). Further, 10 ng of the sulfated-  
20 fucose-containing polysaccharide-F, 10 ng of dextran sulfate powder, or 1  $\mu$ g of sodium alginate was respectively added to the above reaction mixture.

The reaction in 25 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 66°C, 15 seconds. Thereafter, 5  $\mu$ l of the resulting reaction mixture  
25 was subjected to electrophoresis on 1% agarose gel containing ethidium bromide

in an amount of 0.00005%, whereby an amplified fragment was confirmed. The results are shown in Table 2.

Table 2

Acidic Substance	Amount Added	Amplification Results
Sulfated-fucose-containing Polysaccharide-F	10 ng	++
Dextran Sulfate Powder	10 ng	+
Sodium Alginate	1 $\mu$ g	+
No Addition		-

- ++: Intensive amplification being observed;  
 +: Amplification being observed;  
 $\pm$ : Slight amplification being observed; and  
 -: No amplification being observed.

As shown in Table 2, there was confirmed that an expected fragment of 0.5 kb was excellently amplified in any case where acidic substances were added. On the other hand, for a similar reaction mixture without addition of the acidic substances, when PCR was carried out under the above conditions, an amplified fragment of 0.5 kb could not be confirmed.

- (2) Effects of Sulfated-fucose-containing Polysaccharide-F, Sulfated-fucose-containing Polysaccharide-U and Sodium Polyglutamate on DNA Synthesis Reaction by Pfu DNA Polymerase II

Further, studies were conducted on the effects of the sulfated-fucose-containing polysaccharide-F obtained in Example 6 described above, the sulfated-fucose-containing polysaccharide-U obtained in Example 7 described

above, or sodium polyglutamate (manufactured by Sigma) as an acidic substance. A reaction mixture for PCR was prepared in the same manner as described above except for changing the amount of template  $\lambda$ DNA to 500 pg and the primer pairs to primers  $\lambda$ 1 and  $\lambda$ 3. Five nanograms of the sulfated-fucose-containing polysaccharide-F, 5 ng, 10 ng, 20 ng or 30 ng each of the sulfated-fucose-  
5 containing polysaccharide-U, or 250 ng, 500 ng, 750 ng or 1000 ng each of sodium polyglutamate was respectively added to the above reaction mixture.

The reaction was carried out in 30 cycles, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 66°C, 15 seconds. After the  
10 termination of the reaction, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. The results are shown in Table 3.



Table 3

Acidic Substance	Amount Added (ng)	Amplification Results
Sulfated-fucose-containing Polysaccharide-F	5	++
Sulfated-fucose-containing Polysaccharide-U	5	++
	10	+
	20	+
	30	±
Sodium Polyglutamate	250	+
	500	+
	750	++
	1000	+
No Addition		—

- ++: Intensive amplification being observed;  
 +: Amplification being observed;  
 ±: Slight amplification being observed; and  
 -: No amplification being observed.

As shown in Table 3, when the sulfated-fucose-containing polysaccharide-F, the sulfated-fucose-containing polysaccharide-U, or sodium polyglutamate was added, an amplified fragment of 1 kb was confirmed in any of these cases. On the other hand, an amplified fragment of 1 kb could not be confirmed for one without addition of any of these acidic substances.

#### Example 9: Enhancement of DNA Synthesis Reaction of Taq DNA Polymerase by Acidic Substances

Using the sulfated-fucose-containing polysaccharide-F obtained in Example 6 described above or sodium alginate, the effects of these acidic

substances imparted on TaKaRa Taq DNA Polymerase (manufactured by Takara Shuzo Co., Ltd.) were examined. In this example, PCR was carried out in normal mode by using TaKaRa PCR Thermal Cycler Personal.

A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda$ 1 and  $\lambda$ 3 as a primer pair, and TaKaRa Taq DNA Polymerase as DNA polymerase was prepared, and PCR was carried out. The composition of the reaction mixture is as follows.

Composition of Reaction Mixture:

TaKaRa Taq DNA Polymerase buffer, 0.2 mM each of dATP, dCTP, dGTP and dTTP, 1.25 U of TaKaRa Taq DNA Polymerase, 100 pg of  $\lambda$ DNA, and 10 pmol each of primers  $\lambda$ 1 and  $\lambda$ 3 (a final volume being 50  $\mu$ l). Further, 0.1 ng or 0.25 ng each of the sulfated-fucose-containing polysaccharide-F or 1  $\mu$ g of sodium alginate was respectively added to the above reaction mixture.

The reaction was carried out in 30 cycles, wherein one cycle of reaction comprises a process consisting of 98°C, 10 seconds - 68°C, 1 minute. After the termination of the reaction, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. The results are shown in Table 4.

Table 4

Acidic Substance	Amount Added	Amplification Results
Sulfated-fucose-containing Polysaccharide-F	0.1 ng 0.25 ng	++ ++
Sodium Alginate	1 µg	+
No Addition		-

++: Intensive amplification being observed;  
 +: Amplification being observed;  
 ±: Slight amplification being observed; and  
 -: No amplification being observed.

As shown in Table 4, there was confirmed that an expected fragment of 1 kb was excellently amplified in any case where acidic substances were added. On the other hand, for a similar reaction mixture without addition of the acidic substances, when PCR was carried out under the above reaction conditions, an amplified fragment of 1 kb could not be confirmed.

#### Example 10: Effects of Acidic Substances on DNA Polymerase for LA-PCR

(1) The effects of the acidic substances were studied on LA-PCR method comprising a combination of DNA polymerase having 3' → 5' exonuclease activity, and DNA polymerase having no such activity. A reaction mixture comprising λDNA as a template, primers λ1 and λ8 as a primer pair was prepared, and PCR was carried out. In this PCR, TaKaRa EX-Taq DNA Polymerase (manufactured by Takara Shuzo Co., Ltd.) was used. The composition of the reaction mixture is as follows.

In the following example, PCR was carried out in Fast mode by using TaKaRa PCR Thermal Cycler Personal (manufactured by Takara Shuzo Co., Ltd.).

Composition of Reaction Mixture:

5 a reaction mixture comprising TaKaRa EX-Taq DNA Polymerase buffer, 0.2 mM each of dATP, dCTP, dGTP and dTTP, 10 pg of  $\lambda$ DNA, 1.25 U of TaKaRa EX-Taq DNA Polymerase, and 10 pmol each of primers  $\lambda$ 1 and  $\lambda$ 8 (a final volume being 50  $\mu$ l). Further, 1 ng of the sulfated-fucose-containing polysaccharide-F or 0.5  $\mu$ g of sodium alginate was respectively added to the  
10 above reaction mixture.

The reaction was carried out in 27 cycles, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 3 minutes. After the termination of the reaction, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount  
15 of 0.00005%, whereby an amplified fragment was confirmed. The results are shown in Table 5.

Further, PCR was carried out for a reaction having the same composition as the above reaction mixture except for using 100 pg of  $\lambda$ DNA as template DNA, and adding 0.25  $\mu$ g of sodium alginate or 1 ng of the sulfated-fucose-  
20 containing polysaccharide-F as an acidic substance.

The reaction was carried out in 29 cycles, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 3 minutes. After the termination of the reaction, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount  
25 of 0.00005%, whereby an amplified fragment was confirmed. The results are

also shown in Table 5.

Table 5

Amount of Template DNA	Acidic Substance	Amount Added	Amplification Results
10 pg	Sulfated-fucose-containing Polysaccharide-F	1 ng	++
	Sodium Alginate	0.5 $\mu$ g	++
	No Addition	—	—
100 pg	Sulfated-fucose-containing Polysaccharide-F	1 ng	++
	Sodium Alginate	0.25 $\mu$ g	++
	No Addition	—	$\pm$

5

++: Intensive amplification being observed;  
 +: Amplification being observed;  
 $\pm$ : Slight amplification being observed; and  
 -: No amplification being observed.

10

As shown in Table 5, there was confirmed that a fragment of 10 kb when adding an acidic substance was excellently amplified for any of the amounts of template DNA. On the other hand, when PCR was carried out under the above reaction conditions for a reaction mixture without addition of the acidic substances, an amplified fragment of 10 kb could be confirmed very slightly for the case where template DNA was 100 pg.

15

(2) The effects of the acidic substances were studied for KOD dash DNA Polymerase (manufactured by TOYOBO CO., LTD.), comprising a combination

of DNA polymerase having 3' → 5' exonuclease activity, and DNA polymerase having no such activity. A reaction mixture comprising λDNA as a template, primers λ1 and λ9 as a primer pair was prepared, and PCR was carried out. In the following example, PCR was carried out in Fast mode by using TaKaRa PCR Thermal Cycler Personal (manufactured by Takara Shuzo Co., Ltd.). The composition of the reaction mixture is as follows.

Composition of Reaction Mixture:

exclusive buffer attached for KOD dash DNA Polymerase, 0.2 mM each of dATP, dCTP, dGTP and dTTP, 10 pg of λDNA, 2.5 U of KOD dash DNA Polymerase, and 10 pmol each of primers λ1 and λ9 (a final volume being 50 μl). Further, 1 ng of the sulfated-fucose-containing polysaccharide-F or 0.5 μg of sodium alginate was respectively added to the above reaction mixture.

The reaction was carried out in 25 cycles, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 3 minutes. After the termination of the reaction, 5 μl of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. The results are shown in Table 6.

Table 6

Acidic Substance	Amount Added	Amplification Results
Sulfated-fucose-containing Polysaccharide-F	1 ng	+
Sodium Alginate	0.5 $\mu$ g	++
No Addition		$\pm$

- ++: Intensive amplification being observed;  
 +: Amplification being observed;  
 $\pm$ : Slight amplification being observed; and  
 -: No amplification being observed.

As shown in Table 6, there was confirmed that an expected fragment of 12 kb was excellently amplified in any case where acidic substances were added. On the other hand, for a similar reaction mixture without addition of the acidic substances, when PCR was carried out under the above reaction conditions, an amplified fragment of 12 kb could be only slightly confirmed.

#### Example 11: PCR Using Two Kinds of DNA Polymerases Each Having 3' $\rightarrow$ 5'

##### Exonuclease Activity

Studies were conducted on PCR simultaneously using Pfu DNA Polymerase II and any one of KOD DNA Polymerase (manufactured by TOYOBO CO., LTD.), VENT DNA Polymerase (manufactured by New England Biolab), or DEEP VENT DNA Polymerase (manufactured by New England Biolab), each of which is DNA polymerase having 3'  $\rightarrow$  5' exonuclease activity.

A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda$ 1 and  $\lambda$ 8 as a primer pair was prepared, and PCR was carried out. In this PCR, a combination of Pfu DNA Polymerase II and KOD DNA Polymerase, a combination of Pfu DNA Polymerase II and VENT DNA Polymerase, or a combination of Pfu DNA Polymerase II and DEEP VENT DNA Polymerase was respectively used. The composition of the reaction mixture is as follows.

Composition of Reaction Mixture:

10 mM Tris-hydrochloric acid (pH 9.2), 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 500 pg of  $\lambda$ DNA, and 5 pmol each of primers  $\lambda$ 1 and  $\lambda$ 8. To the above reaction mixture was respectively added 0.375 U of Pfu DNA Polymerase II, and 3.75 mU of KOD DNA Polymerase, 6.25 mU of VENT DNA Polymerase or DEEP VENT DNA Polymerase, the reaction mixture making up a final volume of 25  $\mu$ l.

In addition, as controls, a reaction mixture in which Pfu DNA Polymerase II, KOD DNA Polymerase, VENT DNA Polymerase or DEEP VENT DNA Polymerase was added alone was prepared. The reaction in 30 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 3 minutes. Thereafter, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified DNA fragment was confirmed. The results are shown in Table 7.



Table 7

Combination of DNA Polymerase (Polymerase A/Polymerase B)	Amount of Enzyme Used (Polymerase A/ Polymerase B)	Amplification Results
Pfu DNA Polymerase II/ –	0.375 U/ –	±
– /KOD DNA Polymerase	– /3.75 mU	–
– /VENT DNA Polymerase	– /6.25 mU	–
– /DEEP VENT DNA Polymerase	– /6.25 mU	–
Pfu DNA Polymerase II/ KOD DNA Polymerase	0.375 U/3.75 mU	+
Pfu DNA Polymerase II/ VENT DNA Polymerase	0.375 U/6.25 mU	+
Pfu DNA Polymerase II/ DEEP VENT DNA Polymerase	0.375 U/6.25 mU	+

++: Intensive amplification being observed;  
 +: Amplification being observed;  
 ±: Slight amplification being observed; and  
 -: No amplification being observed.

5

As shown in Table 7, while an amplified fragment of 10 kb was not found when using KOD DNA Polymerase, VENT DNA Polymerase or DEEP VENT DNA Polymerase alone, there was found a slight amount of an amplified fragment of 10 kb when using Pfu DNA Polymerase II alone. Further, in a system where 0.375 U of Pfu DNA Polymerase II, and 3.75 mU of KOD DNA Polymerase, 6.25 mU of VENT DNA Polymerase or DEEP VENT DNA Polymerase is used in combination, there was confirmed that an amplification efficiency of a fragment of 10 kb was improved, as compared with the case of using Pfu DNA Polymerase II alone.

15

### Example 12

PCR was carried out by using in combination Pfu DNA Polymerase II obtained in Example 1 and Pfu DNA Polymerase I (manufactured by Stratagene), which is  $\alpha$ -type DNA polymerase, and sodium alginate, which is an acidic substance.

A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda$ 1 and  $\lambda$ 3 as a primer pair was prepared, and PCR was carried out. The composition of the reaction mixture is as follows.

#### Composition of Reaction Mixture:

reaction buffer for PCR (10 mM Tris-hydrochloric acid, pH 9.2, 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, and 0.004% of sodium alginate), 10 pg of  $\lambda$ DNA, and 5 pmol each of primers  $\lambda$ 1 and  $\lambda$ 3. To the above reaction mixture was added an enzyme solution containing 1.25 U of Pfu DNA Polymerase II and 1.14 U of Pfu DNA Polymerase I, the reaction mixture making up a final volume of 25  $\mu$ l.

In addition, a reaction mixture without addition of sodium alginate was prepared as a control. The reaction in 30 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 66°C, 15 seconds. Thereafter, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified DNA fragment was confirmed. The results are shown in Table 8.

Table 8

Kinds of Kit Used	Amplification Results
Reaction Buffer (Containing 0.004% Sodium Alginate)	++
Reaction Buffer (Without Containing Sodium Alginate)	—

++: Intensive amplification being observed;  
 +: Amplification being observed;  
 ±: Slight amplification being observed; and  
 -: No amplification being observed.

5

As shown in Table 8, there was confirmed that amplification efficiency for a fragment of 1 kb was improved in the reaction in which Pfu DNA Polymerase I, Pfu DNA Polymerase II and sodium alginate were used in combination. Further, the above study has been conducted for commercially available sodium alginate for each viscosity (100 to 1000 centipoises), and as a result, completely the same improvements in the amplification efficiency for DNA synthesis were confirmed.

10

15

#### Example 13: Preparation of Kit

A PCR kit (20 reactions) of the present invention was constructed by using Pfu DNA Polymerase II obtained in Example 1, Pfu DNA Polymerase I and sodium alginate, which is an acidic substance in combination.

20

The composition for the kit is shown below:

10 × Reaction Buffer for PCR	50 µl
100 mM Tris-Hydrochloric Acid (pH 9.2)	
750 mM Potassium Chloride	
0.1% BSA	
0.04% Sodium Alginate (100 to 150 centipoises)	
25 mM Magnesium Chloride Solution	120 µl
2.5 mM dNTP Mix (2.5 mM Each of dATP, dCTP, dGTP and dTTP)	80 µl
Enzyme Mixed Solution for DNA Polymerases (2.5 U of Pfu DNA Polymerase II and 2.28 U of Pfu DNA Polymerase I / 1 µl)	10 µl

A reaction mixture for PCR was prepared using the above kit.  $\lambda$ DNA was used as a template. The composition for the reaction mixture is shown in Table 9.

5

Table 9

Composition of Reaction Mixture	
10 × PCR Reaction Buffer	2.5 µl
Magnesium Chloride Solution	6 µl
dNTP Mix	4 µl
Enzyme Mixed Solution for DNA Polymerase	0.5 µl
$\lambda$ DNA	10 pg
$\lambda$ 1 Primer	5 pmol
$\lambda$ 3 Primer	5 pmol
Sterilized Distilled Water	Balance
Final Volume	25 µl

In addition, a reaction mixture having the same composition as described above except that sodium alginate was not contained was prepared as a control.

The reaction for the above reaction mixture was carried out under the PCR conditions shown in Example 12. As a result, there could be confirmed that the reaction mixture prepared by using the kit containing sodium alginate described above had improved amplification efficiency of a fragment of 1 kb, as compared to that of control.

#### Example 14: Numerical Explanation on Enhancement of DNA Synthesis

##### Reaction by Acidic Substance

Using the sulfated-fucose-containing polysaccharide-F obtained in Example 6 described above, the effects on the activity of Pfu DNA Polymerase II were examined. A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda$ 1 and  $\lambda$ 2 as a primer pair, and Pfu DNA Polymerase II as DNA polymerase was prepared, and PCR was carried out. The composition of the reaction mixture is as follows.

##### Composition of Reaction Mixture:

10 mM Tris-hydrochloric acid, pH 9.2, 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 1.25 U of Pfu DNA Polymerase II, 500 pg of  $\lambda$ DNA, and 5 pmol each of primers  $\lambda$ 1 and  $\lambda$ 2 (a final volume being 25  $\mu$ l). Further, 10 ng of the sulfated-fucose-containing polysaccharide-F was respectively added to the above reaction mixture.

The reaction in 30 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C,

5 seconds - 66°C, 15 seconds. Thereafter, 5 µl of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an expected amplified fragment of 0.5 kb was confirmed. In addition, the agarose gel was analyzed by using a fluorescent image analyzer FM-BIO (manufactured by Takara Shuzo Co., Ltd.) to numerically explain the relative amount of the amplified fragment.

As a result, as compared with a case where a reaction system in which an acidic substance was not added was considered to be 1, one using sulfated-fucose-containing polysaccharide-F, which is an acidic substance, was 4.4. In other words, there could be confirmed that an amount of amplified product was multiplied by 4.4 by the addition of an acidic substance in PCR.

#### Example 15: Enhancement of DNA Synthesis Reaction by Acidic Substance

Using a polyacrylic acid as an acidic substance, the effects on the activity of Pfu DNA Polymerase II were examined. A reaction mixture comprising λDNA as a template, primers λ1 and λ3 as a primer pair, and Pfu DNA Polymerase II as DNA polymerase was prepared, and PCR was carried out. The composition of the reaction mixture is as follows.

#### Composition of Reaction Mixture:

10 mM Tris-hydrochloric acid, pH 9.2, 75 mM potassium chloride, 6 mM magnesium chloride, 0.4 mM each of dATP, dCTP, dGTP and dTTP, 0.01% BSA, 1.25 U of Pfu DNA Polymerase II, 500 pg of λDNA, and 5 pmol each of primers λ1 and λ3 (a final volume being 25 µl). Further, 100 ng of a polyacrylic acid having an average molecular weight of about 5,000, about 25,000, about 250,000, or about 1,000,000 (each being manufactured by Wako Pure

Chemicals) was respectively added to the above reaction mixture.

The reaction in 30 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 66°C, 15 seconds. Thereafter, 5 µl of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. The results are shown in Table 10.

Table 10

Acidic Substance	Average Molecular Weight	Amplification Results
Sodium Polyacrylate	5,000	++
	25,000	++
	250,000	++
	1,000,000	++
No Addition		-

- ++: Intensive amplification being observed;  
 +: Amplification being observed;  
 ±: Slight amplification being observed; and  
 -: No amplification being observed.

As shown in Table 10, there was confirmed that an expected fragment of 1 kb was excellently amplified in any case where the sodium polyacrylate of any of the average molecular weights was added. On the other hand, for a similar reaction mixture without addition of the acidic substance, when PCR was carried out under the above reaction conditions, an amplified fragment of 1 kb could not

be confirmed.

Example 16: Effects of Acidic Substances on  $\alpha$ -Type DNA Polymerase

(1) Using sodium alginate, sodium hyaluronate (manufactured by Seikagaku Kogyo), or  $\kappa$  carrageenan (manufactured by Sigma), the effects of these acidic substances on  $\alpha$ -type DNA polymerase were examined. A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda$ 1 and  $\lambda$ 8 as a primer pair, and KOD DNA Polymerase (manufactured by TOYOBO CO., LTD.) as DNA polymerase was prepared, and PCR was carried out. The composition of the reaction mixture is as follows.

Composition of Reaction Mixture:

KOD DNA Polymerase Buffer I (manufactured by TOYOBO CO., LTD.)  
0.2 mM each of dATP, dCTP, dGTP and dTTP, 0.625 U of KOD DNA Polymerase, 100 pg of  $\lambda$ DNA, and 5 pmol each of primers  $\lambda$ 1 and  $\lambda$ 8 (a final volume being 25  $\mu$ l). Further, 1  $\mu$ g or 2.5  $\mu$ g of sodium alginate, 0.1  $\mu$ g or 0.5  $\mu$ g of sodium hyaluronate, or 0.1  $\mu$ g or 0.25  $\mu$ g of  $\kappa$  carrageenan was respectively added to the above reaction mixture.

The reaction in 30 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 3 minutes. Thereafter, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 1% agarose gel containing ethidium bromide in an amount of 0.00005%, whereby an amplified fragment was confirmed. The results are shown in Table 11.



Table 11

Acidic Substance	Amount Added ( $\mu$ g)	Amplification Results
Sodium Alginate	1	++
	2.5	++
Sodium Hyaluronate	0.1	+
	0.5	+
$\kappa$ Carrageenan	0.1	+
	0.25	+
No Addition		$\pm$

- ++: Intensive amplification being observed;  
 +: Amplification being observed;  
 $\pm$ : Slight amplification being observed; and  
 -: No amplification being observed.

As shown in Table 11, there was confirmed that an expected fragment of 10 kb was excellently amplified in any case where acidic substances were added. On the other hand, for a similar reaction mixture without addition of the acidic substances, when PCR was carried out under the above reaction conditions, an amplified fragment of 10 kb could be confirmed weakly.

(2) The effects on  $\alpha$ -type DNA polymerase were also examined in the same manner as in item (1) above for sodium alginate,  $\kappa$  carrageenan, sodium heparan sulfate (manufactured by Sigma), sodium dermatan sulfate (manufactured by Sigma), potassium polyvinyl sulfate (manufactured by nacalaitesque) or sodium polystyrenesulfonate (two kinds: average molecular weights of 5,400 and 35,000, all manufactured by nacalaitesque). A reaction mixture comprising  $\lambda$ DNA as a template, primers  $\lambda$ 1 and  $\lambda$ 8 as a primer pair, and KOD DNA Polymerase as

DNA polymerase was prepared, and PCR was carried out. The composition of the reaction mixture is as follows.

Composition of Reaction Mixture:

KOD DNA Polymerase Buffer II (manufactured by TOYOBO CO., LTD.)

5 0.2 mM each of dATP, dCTP, dGTP and dTTP, 1.25 U of KOD DNA  
Polymerase, 100 pg of  $\lambda$ DNA, and 10 pmol each of primers  $\lambda$ 1 and  $\lambda$ 8 (a final  
volume being 50  $\mu$ l). Further, 2.5  $\mu$ g or 5  $\mu$ g of sodium alginate, 0.25  $\mu$ g or  
0.5  $\mu$ g of  $\kappa$  carrageenan, or 125 ng, 250 ng or 375 ng of sodium dermatan sulfate,  
10 500 ng of sodium heparan sulfate, 10 ng of potassium polyvinyl sulfate, 2.5 ng or  
5 ng of sodium polystyrenesulfonate (average molecular weight: 5,400), or 5 ng  
of sodium polystyrenesulfonate (average molecular weight: 35,000) was  
respectively added to the above reaction mixture.

The reaction in 30 cycles was carried out for each of these reaction  
mixtures, wherein one cycle of reaction comprises a process consisting of 98°C,  
15 5 seconds - 68°C, 3 minutes. Thereafter, 5  $\mu$ l of the resulting reaction mixture  
was subjected to electrophoresis on 1% agarose gel containing ethidium bromide  
in an amount of 0.00005%, whereby an amplified fragment was confirmed. The  
results are shown in Table 12.

Table 12

Acidic Substance	Amount Added	Amplification Results
Sodium Alginate	2.5 µg	++
	5 µg	++
κ Carrageenan	0.25 µg	++
	0.5 µg	++
Sodium Dermatan Sulfate	125 ng	+
	250 ng	+
	375 ng	+
Sodium Heparan Sulfate	500 ng	+
Potassium Polyvinyl Sulfate	10 ng	+
Sodium Polystyrenesulfonate (Average Molecular Weight: 5,400) (Average Molecular Weight: 35,000)	2.5 ng	+
	5 ng	+
	5 ng	+
No Addition		—

- ++: Intensive amplification being observed;  
 +: Amplification being observed;  
 ±: Slight amplification being observed; and  
 -: No amplification being observed.

5

As shown in Table 12, there was confirmed that an expected fragment of 10 kb was excellently amplified in any case where acidic substances were added. On the other hand, for a similar reaction mixture without addition of the acidic substances, when PCR was carried out under the above reaction conditions, an amplified fragment of 10 kb could not be confirmed.

10

#### Example 17: Effects of Cationic Complexes on Various Kinds of DNA

15

#### Polymerases

Using hexaamminecobalt (III) chloride ( $[\text{Co}(\text{NH}_3)_6]\text{Cl}_3$ ) (manufactured by Sigma), tris(ethylenediamine)cobalt (III) chloride ( $[\text{Co}(\text{C}_2\text{H}_8\text{N}_2)_3]\text{Cl}_3$ ) (manufactured by Aldrich), or tris(ethylenediamine)rhodium (III) trichloride•trihydrate ( $[\text{Rh}(\text{C}_2\text{H}_8\text{N}_2)_3]\text{Cl}_3 \cdot 3\text{H}_2\text{O}$ ) (manufactured by Aldrich), the effects on the amplification reaction by various kinds of DNA polymerases were examined. As the DNA polymerases, TaKaRa Taq DNA Polymerase (hereinafter referred to as “Taq DNA Polymerase” manufactured by Takara Shuzo Co., Ltd.), Pyrobest DNA Polymerase (manufactured by Takara Shuzo Co., Ltd.) and Ex Taq DNA Polymerase (manufactured by Takara Shuzo Co., Ltd.) were used.

A reaction mixture comprising *Escherichia coli* JM109 genomic DNA as a template, primers Eco1 and Eco2 as a primer pair, and DNA polymerase was prepared, and PCR was carried out. The compositions of the reaction mixture for each enzyme are as follows.

#### Compositions of Reaction Mixtures:

##### 1) Reaction System for Taq DNA Polymerase

5  $\mu\text{l}$  of buffer for  $10 \times$  Taq PCR buffer (manufactured by Takara Shuzo Co., Ltd.), 0.2 mM each of dATP, dCTP, dGTP and dTTP, 1.25 U of Taq DNA Polymerase, 100 pg of *Escherichia coli* JM109 genomic DNA, and 10 pmol each of primers Eco1 and Eco2 (a final volume being 50  $\mu\text{l}$ ).

##### 2) Reaction System for Pyrobest DNA Polymerase

5  $\mu\text{l}$  of 10-fold concentrated Pyrobest buffer (manufactured by Takara Shuzo Co., Ltd.), 0.2 mM each of dATP, dCTP, dGTP and dTTP, 1.25 U of Pyrobest DNA Polymerase, 100 pg of *Escherichia coli* JM109 genomic DNA,

and 10 pmol each of primers Eco1 and Eco2 (a final volume being 50  $\mu$ l).

### 3) Reaction System for ExTaq DNA Polymerase

a reaction mixture comprising 5  $\mu$ l of 10-fold concentrated buffer for TaKaRa ExTaq DNA Polymerase (manufactured by Takara Shuzo Co., Ltd.), 0.2 mM each of dATP, dCTP, dGTP and dTTP, 100 pg of *Escherichia coli* JM109 genomic DNA, 1.25 U of TaKaRa EX-Taq DNA Polymerase, and 10 pmol each of primers Eco1 and Eco2 (a final volume being 50  $\mu$ l).

Further, an aqueous solution of the above-mentioned complexes each so as to have a final concentration of 50  $\mu$ M, 100  $\mu$ M, or 200  $\mu$ M was added to the above reaction mixture.

The reaction was carried out in Fast mode by using TaKaRa PCR Thermal Cycler Personal (manufactured by Takara Shuzo Co., Ltd.). The reaction conditions are as follows. When using Taq DNA Polymerase and Pyrobest DNA Polymerase, the reaction in 30 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 2 minutes, and when using ExTaq DNA Polymerase, the reaction in 30 cycles was carried out for each of these reaction mixtures, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 1 minute. After the termination of the reaction, 5  $\mu$ l of the resulting reaction mixture was subjected to electrophoresis on 2% agarose gel containing ethidium bromide in an amount of 0.00005%. The gel after the electrophoresis was subjected to ultraviolet irradiation to visualize the band ascribed to the amplified product, whereby an amplified fragment was confirmed. The amount of

amplified DNA was numerically explained by using a fluorescent image analyzer FM-BIO 100 (manufactured by Takara Shuzo Co., Ltd.). The results are shown in Table 13.

5

Table 13

Cationic Complex	DNA Polymerase			
	Taq	Pyrobest	ExTaq	
$[\text{Co}(\text{NH}_3)_6]\text{Cl}_3$				
50 $\mu\text{M}$	2.3	2.1	1.9	
100 $\mu\text{M}$	2.9	2.1	1.9	
200 $\mu\text{M}$	—	2.1	—	
$[\text{Co}(\text{H}_2\text{NCH}_2\text{CH}_2\text{NH}_2)_3]\text{Cl}_3$				
50 $\mu\text{M}$	2.6	2.1	2.0	
100 $\mu\text{M}$	3.0	2.2	2.0	
200 $\mu\text{M}$	—	—	—	
$[\text{Rh}(\text{H}_2\text{NCH}_2\text{CH}_2\text{NH}_2)_3]\text{Cl}_3$				
50 $\mu\text{M}$	1.4	1.4	1.3	
100 $\mu\text{M}$	2.1	1.7	1.5	
200 $\mu\text{M}$	2.5	2.0	1.7	
No Addition	1	1	1	

10

As a result, with regard to Taq DNA Polymerase, there was confirmed an increase in the amount of DNA amplified in about 1.4 times to about 3 times that of a case of no addition, in the reaction in which 50  $\mu\text{M}$  or 100  $\mu\text{M}$  of hexaamminecobalt (III) chloride, 50  $\mu\text{M}$  or 100  $\mu\text{M}$  of tris(ethylenediamine)cobalt (III) chloride, or 50  $\mu\text{M}$ , 100  $\mu\text{M}$  or 200  $\mu\text{M}$  of

tris(ethylenediamine)rhodium (III) trichloride was added.

Next, with regard to Pyrobest DNA Polymerase, there was confirmed an increase in the amount of DNA amplified in about 1.4 times to about 2.2 times that of a case of no addition, in the reaction in which 50  $\mu$ M, 100  $\mu$ M or 200  $\mu$ M of  
 5 of hexaamminecobalt (III) chloride, 50  $\mu$ M or 100  $\mu$ M of tris(ethylenediamine)cobalt (III) chloride, or 50  $\mu$ M, 100  $\mu$ M or 200  $\mu$ M of tris(ethylenediamine)rhodium (III) trichloride was added.

Further, with regard to ExTaq DNA Polymerase, there was confirmed an increase in the amount of DNA amplified in about 1.3 times to about 2 times that  
 10 of a case of no addition, in the reaction in which 50  $\mu$ M or 100  $\mu$ M of hexaamminecobalt (III) chloride, 50  $\mu$ M or 100  $\mu$ M of tris(ethylenediamine)cobalt (III) chloride, or 50  $\mu$ M, 100  $\mu$ M or 200  $\mu$ M of tris(ethylenediamine)rhodium (III) trichloride was added.

#### 15 Example 18: Effects on Reaction Time by Addition of Hexaamminecobalt (III) Chloride

With regard to each DNA polymerase, the effects on shortening the reaction time by the addition of hexaamminecobalt (III) chloride were examined.

The DNA polymerases used and the composition for the reaction mixture  
 20 were similar to those in Example 17, except for the cationic complex, where in cases of using Taq DNA Polymerase and Pyrobest DNA Polymerase, hexaamminecobalt (III) chloride was added so as to have a final concentration of 100  $\mu$ M, and in a case of using ExTaq DNA Polymerase, hexaamminecobalt (III) chloride was added so as to have a final concentration of 50  $\mu$ M, respectively.

25 The reaction was carried out in Fast mode by using TaKaRa PCR Thermal

Cycler Personal. The reaction conditions are as follows. When using Taq DNA Polymerase and Pyrobest DNA Polymerase, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 60 seconds or 80 seconds or 100 seconds or 120 seconds.

5 When using ExTaq DNA Polymerase, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 30 seconds or 40 seconds or 50 seconds or 60 seconds.

10 On the other hand, as a control, the time period or number of cycles for annealing and extension reaction were determined so as to obtain the same amount of the amount of amplified product for one without addition of the cationic complex mentioned above.

15 After the termination of the reaction, 5 µl of the resulting reaction mixture was subjected to electrophoresis on 2% agarose gel containing ethidium bromide in an amount of 0.00005%. The gel after the electrophoresis was subjected to ultraviolet irradiation to visualize the band ascribed to the amplified product, whereby an amplified fragment was confirmed. The results are shown in Table 14.



Table 14

DNA Polymerase	Cationic Complex	
	Addition	No Addition
Taq and Pyrobest	98°C, 5 s - 68°C, 60 s 30 Cycles (Time Period Required: 2953 s)	98°C, 5 s - 68°C, 120 s 31 Cycles (Time Period Required: 4911 s)
ExTaq	98°C, 5 s - 68°C, 40 s 30 Cycles (Time Period Required: 2294 s)	98°C, 5 s - 60°C, 60 s 30 Cycles (Time Period Required: 2953 s)

As shown in Table 14, in the cases of using Taq DNA Polymerase and Pyrobest DNA polymerase, when hexaamminecobalt (III) chloride was added in an amount of 100  $\mu$ M, the reaction time period could be shortened to about 3/5 that of the case of no addition. Further, in the case of using ExTaq DNA Polymerase, when hexaamminecobalt (III) chloride was added in an amount of 50  $\mu$ M, the reaction time period could be shortened to about 4/5 that of the case of no addition.

#### Example 19: Effects on Amplification Sensitivity by Addition of Hexaamminecobalt (III) Chloride

With regard to each of DNA polymerases, the effects on the amplification sensitivity by the addition of hexaamminecobalt (III) chloride were examined.

The DNA polymerases used and the composition for the reaction mixture were similar to those in Example 17, except for the cationic complex, where in cases of using Taq DNA Polymerase and Pyrobest DNA Polymerase, hexaamminecobalt (III) chloride was added so as to have a final concentration of

100  $\mu$ M, or in a case of using ExTaq DNA Polymerase, hexaamminecobalt (III) chloride was added so as to have a final concentration of 50  $\mu$ M, respectively. Further, the amount of template DNA was added so as to be 1 pg, 5 pg, 10 pg, 100 pg, or 1 ng, respectively.

5           The reaction was carried out in Fast mode by using TaKaRa PCR Thermal Cycler Personal. The reaction conditions are as follows. When using Taq DNA Polymerase and Pyrobest DNA Polymerase, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 2 minutes. When using ExTaq DNA Polymerase, the reaction  
10 in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 1 minute.

On the other hand, as a control, the reaction for one without addition of the cationic complex was carried out in the same manner.

15           After the termination of the reaction, 5  $\mu$ l of each of the resulting reaction mixtures was subjected to electrophoresis on 2% agarose gel containing ethidium bromide in an amount of 0.00005%. The gel after the electrophoresis was subjected to ultraviolet irradiation to visualize the band ascribed to the amplified product, whereby an amplified fragment was confirmed.

20           As a result, there could be confirmed that in any case where any enzymes were used, the addition of hexaamminecobalt (III) chloride allowed to amplify even in an amount of template DNA of 1 pg, amplification can be carried out by. On the other hand, in the case of no addition, amplification could not be carried out just with an amount of template DNA of 1 pg.

#### 25           Example 20: Effects on Amplification Reaction in Low-Magnesium

### Concentration Reaction Mixture by Addition of Hexaamminecobalt (III)

#### Chloride

Using hexaamminecobalt (III) chloride, the effects on the amplification reaction in a reaction mixture having a magnesium concentration lower than that of a usually employed one were examined.

The DNA polymerases used and the composition for the reaction mixture were similar to those in Example 17, except for using each of magnesium-free, 10 × PCR buffers (each being manufactured by Takara Shuzo Co., Ltd.) for each DNA polymerase. Further, with regard to magnesium chloride, magnesium chloride was added so as to have an amount smaller than the usual content for each of Taq DNA Polymerase (the amount being 0.75 mM against the usual amount of 1.5 mM), Pyrobest DNA Polymerase (the amount being 0.5 mM against the usual amount of 1 mM), or ExTaq DNA Polymerase (the amount being 1.25 mM against the usual amount of 2 mM). Further, with regard to the cationic complex, in cases of using Taq DNA Polymerase and Pyrobest DNA Polymerase, hexaamminecobalt (III) chloride was added so as to have a final concentration of 100 μM, or in a case of using ExTaq DNA Polymerase, hexaamminecobalt (III) chloride was added so as to have a final concentration of 10 μM, respectively.

The reaction was carried out in Fast mode by using TaKaRa PCR Thermal Cycler Personal. The reaction conditions are as follows. When using Taq DNA Polymerase and Pyrobest DNA Polymerase, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 2 minutes. When using ExTaq DNA Polymerase, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process

consisting of 98°C, 5 seconds - 68°C, 1 minute.

On the other hand, the reaction for one without addition of the cationic complex was carried out in the same manner as a control.

After the termination of the reaction, 5 µl of the resulting reaction mixture was subjected to electrophoresis on 2% agarose gel containing ethidium bromide in an amount of 0.00005%. The gel after the electrophoresis was subjected to ultraviolet irradiation to visualize the band ascribed to the amplified product, whereby an amplified fragment was confirmed.

As a result, there was confirmed that in all of DNA polymerases, the addition of the cationic complex allowed to amplify in a case where the magnesium concentration was made smaller than the usual content. However, in the case of no addition of cationic complex, amplification could not be carried out.

#### 15 Example 21: Effects of Low-Primer Concentration in Reaction Mixture on Amplification Reaction by Addition of Hexaamminecobalt (III) Chloride

Using hexaamminecobalt (III) chloride, the effects of a reaction mixture having a primer concentration lower than that of a usually employed one on the amplification reaction were examined.

20 The DNA polymerases used and the composition for the reaction mixture were similar to those in Example 17, except for adding two kinds of primers used each in an amount of 2, 2.5, 3.3, 5, 10 or 20 pmol, respectively. Further, with regard to the cationic complex, in cases of using Taq DNA Polymerase and Pyrobest DNA Polymerase, hexaamminecobalt (III) chloride was added so as to  
25 have a final concentration of 100 µM, or in a case of using ExTaq DNA

Polymerase, hexaamminecobalt (III) chloride was added so as to have a final concentration of 50  $\mu$ M, respectively.

The reaction was carried out in Fast mode by using TaKaRa PCR Thermal  
Cycler Personal. The reaction conditions are as follows. When using Taq DNA  
Polymerase and Pyrobest DNA Polymerase, the reaction in 30 cycles was carried  
out, wherein one cycle of reaction comprises a process consisting of 98°C,  
5 seconds - 68°C, 2 minutes. When using ExTaq DNA Polymerase, the reaction  
in 30 cycles was carried out, wherein one cycle of reaction comprises a process  
consisting of 98°C, 5 seconds - 68°C, 1 minute.

On the other hand, the reaction for one without addition of the cationic  
complex was carried out in the same manner as a control.

After the termination of the reaction, 5  $\mu$ l of each of the resulting reaction  
mixtures was subjected to electrophoresis on 2% agarose gel containing ethidium  
bromide in an amount of 0.00005%. The gel after the electrophoresis was  
subjected to ultraviolet irradiation to visualize the band ascribed to the amplified  
product, whereby an amplified fragment was confirmed.

As a result, there was confirmed that in a case of using Taq DNA  
Polymerase, a fragment can be amplified even with a primer amount of as little  
as 2 pmol in the presence of the cationic complex. On the other hand, in the case  
of no addition of the cationic complex, amplification could not be carried out.  
Next, in a case of using Pyrobest DNA Polymerase, a fragment can be amplified  
even with a primer amount of as little as 3.3 pmol in the presence of the cationic  
complex. On the other hand, in the case of no addition of the cationic complex,  
amplification could not be carried out. Further, in a case of using ExTaq DNA  
Polymerase, a fragment can be amplified even with a primer amount of as little

as 2.5 pmol in the presence of the cationic complex. On the other hand, in the case of no addition of the cationic complex, amplification could not be carried out.

5     Example 22: Effects on Enzyme Amount and Amplification Reactions by  
       Addition of Hexaamminecobalt (III) Chloride

      Using hexaamminecobalt (III) chloride, the effects on amplification reaction with an enzyme in an amount of one-half that of usually used.

10     As the enzymes, Taq DNA Polymerase and ExTaq DNA Polymerase were used.

      The composition for the reaction mixture was similar to those in Example 17, except for the amount of DNA polymerase used. In other words, 0.625 U of DNA polymerase was used. Further, with regard to the cationic complex, in cases of using Taq DNA Polymerase and Pyrobest DNA Polymerase,  
 15     hexaamminecobalt (III) chloride was added so as to have a final concentration of 100  $\mu$ M, or in a case of using ExTaq DNA Polymerase, hexaamminecobalt (III) chloride was added so as to have a final concentration of 50  $\mu$ M, respectively.

      The reaction was carried out in Fast mode by using TaKaRa PCR Thermal  
 20     Cycler Personal. The reaction conditions are as follows. When using Taq DNA Polymerase, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 2 minutes. When using ExTaq DNA Polymerase, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C,  
 25     5 seconds - 68°C, 1 minute.

      On the other hand, as a control, the reaction for one using 1.25 U of DNA

polymerase without addition of the cationic complex was carried out in the same manner.

After the termination of the reaction, 5 µl of each of the resulting reaction mixtures was subjected to electrophoresis on 2% agarose gel containing ethidium bromide in an amount of 0.00005%. The gel after the electrophoresis was subjected to ultraviolet irradiation to visualize the band ascribed to the amplified product, whereby an amplified fragment was confirmed.

As a result, there was confirmed that in either case of using Taq DNA Polymerase or ExTaq DNA Polymerase, even if the amount is one-half the usual amount, the fragment can be amplified in the presence of the cationic complex to almost the same level as the usually used amount of the case of no addition.

#### Example 23: Effects on Addition of Cationic Complex When Amplifying GC-Rich Region

The effects on addition of the cationic complex were examined when PCR amplification was carried out with a GC-rich region as a target. As the cationic complex, there was used tris(ethylenediamine)cobalt (III) chloride, or tris(ethylenediamine)rhodium (III) trichloride•trihydrate. Template DNA was prepared from HT29 cells by conventional method. Amplified region and amplified chain lengths were human ApoE genomic region and 441 bp. The GC content of this amplified region was about 74%. As DNA polymerase, TaKaRa LA-Taq DNA Polymerase (manufactured by Takara Shuzo Co., Ltd., hereinafter referred to as "LA-Taq DNA Polymerase") was used. The composition for a reaction mixture is shown below.

Composition for Reaction Mixture:

25  $\mu$ l of  $2 \times$  GC buffer I (manufactured by Takara Shuzo Co., Ltd.), 0.4 mM each of dATP, dCTP, dGTP and dTTP, 2.5 U of LA-Taq DNA Polymerase, 100 ng of HT29 cells genomic region, and 20 pmol each of primers ApoE-1 and ApoE-2 (a final volume being 50  $\mu$ l). Each of the above cationic complex was added to the  
5 above reaction mixture so as to have a final concentration of 50, 100, 200, or 500  $\mu$ M, respectively.

The reaction was carried out in Fast mode by using TaKaRa PCR Thermal  
Cycler Personal. As regarding the reaction conditions, the reaction in 30 cycles  
was carried out, wherein one cycle of reaction comprises a process consisting of  
10 98°C, 5 seconds - 64°C, 10 seconds - 72°C, 40 seconds. On the other hand, as a control, the reaction for one without addition of the cationic complex was carried out in the same manner.

After the termination of the reaction, 5  $\mu$ l of each of the resulting reaction  
mixtures was subjected to electrophoresis on 2% agarose gel containing ethidium  
15 bromide in an amount of 0.00005%. The gel after the electrophoresis was subjected to ultraviolet irradiation to visualize the band ascribed to the amplified product, whereby an amplified fragment was confirmed.

As a result, in a case of adding tris(ethylenediamine)cobalt (III) chloride,  
there was confirmed that the DNA synthesis reaction was enhanced as the  
20 concentration of added complex was increased. In addition, in a case of adding tris(ethylenediamine)rhodium (III) trichloride•trihydrate, there was also confirmed that the DNA synthesis reaction was enhanced as the concentration of added complex was increased as 100  $\mu$ M, 200  $\mu$ M and 500  $\mu$ M. However, in the case of no addition for neither of the cationic complexes, amplification could not  
25 be carried out.



Example 24: Effects on Amplified Chain Length When Adding Cationic Complex

The effects on the amplified chain length when adding a cationic complex were examined. As the cationic complex, there was used hexaamminecobalt (III) trichloride. As template DNA, there was used *Escherichia coli* genome or  $\lambda$ DNA. As primers, in cases of using Taq DNA Polymerase and Pyrobest DNA Polymerase, there was used a combination of Eco1 and Eco5, and in a case of using LA-Taq DNA Polymerase, there was used a combination of  $\lambda$ L36 and  $\lambda$ R485. As DNA polymerase, there was used Taq DNA Polymerase, Pyrobest DNA Polymerase or LA-Taq DNA Polymerase. The composition for the reaction mixture was similar to those in Example 17, except for using the above primer pairs and 10 ng of *Escherichia coli* genome as template DNA in cases of using Taq DNA Polymerase and Pyrobest DNA Polymerase. To the above reaction mixture was added hexaamminecobalt (III) trichloride so as to have a final concentration of 10  $\mu$ M or 20  $\mu$ M in a case of using Taq DNA Polymerase, or was added hexaamminecobalt (III) trichloride so as to have a final concentration of 50  $\mu$ M or 100  $\mu$ M in a case of using Pyrobest DNA Polymerase, respectively. As a control, the one without addition of the cationic complex was also prepared in the same manner.

On the other hand, the composition for a reaction mixture when using LA-Taq DNA Polymerase is shown below.

5  $\mu$ l of 10 $\times$  LA-PCR Buffer II ( $Mg^{2+}$  plus) (manufactured by Takara Shuzo Co., Ltd.), 0.4 mM each of dATP, dCTP, dGTP and dTTP, 2.5 U of LA-Taq DNA Polymerase, 1 ng of  $\lambda$ DNA, and 20 pmol each of primers  $\lambda$ L36 and

$\lambda$ R485 (a final volume being 50  $\mu$ l). Hexaamminecobalt (III) trichloride was added to the above reaction mixture so as to have a final concentration of 50  $\mu$ M. As a control, the one without addition of the cationic complex was also prepared in the same manner.

5           The reaction was carried out in Fast mode by using TaKaRa PCR Thermal Cycler Personal. The reaction conditions are as follows. When using Taq DNA Polymerase and Pyrobest DNA Polymerase, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C, 5 seconds - 68°C, 10 minutes. When using LA-Taq DNA Polymerase, after  
10       treating at 94°C, 1 minute, the reaction in 30 cycles was carried out, wherein one cycle of reaction comprises a process consisting of 98°C, 10 seconds - 68°C, 15 minutes, and thereafter the reaction was completed by treating at 72°C, 15 minutes.

          After the termination of the reaction, 5  $\mu$ l of each of the resulting reaction  
15       mixtures was subjected to electrophoresis on 2% agarose gel containing ethidium bromide in an amount of 0.00005%. The gel after the electrophoresis was subjected to ultraviolet irradiation to visualize the band ascribed to the amplified product, whereby an amplified fragment was confirmed.

          As a result, when using Taq DNA Polymerase, a desired DNA fragment  
20       could be excellently amplified even when hexaamminecobalt (III) trichloride was added at any of concentrations. However, in the case of no addition, a fragment was only slightly amplified. Next, when using Pyrobest DNA Polymerase, a desired DNA fragment could be excellently amplified even when hexaamminecobalt (III) trichloride was added at any of concentrations. However,  
25       in the case of no addition, a fragment was only slightly amplified. Further, when

using LA-Taq DNA Polymerase, a desired DNA fragment could be excellently amplified even when hexaamminecobalt (III) trichloride was added at any of concentrations. However, in the case of no addition, a fragment was only slightly amplified.

5

#### INDUSTRIAL APPLICABILITY

According to the present invention, there is provided a DNA synthesis reaction-enhancer which enhances DNA synthesis reaction by DNA polymerase. The enhancer exhibits an action for various kinds of DNA polymerases, and enhances the DNA synthesis reaction. In addition, the present invention provides a DNA synthesis reaction composition which is capable of carrying out DNA synthesis at high efficiency. The DNA synthesis reaction-enhancer and the DNA synthesis reaction composition of the present invention can be utilized for various processes using DNA polymerases, for instance, PCR method, and can be utilized for research agents for genetic engineering.

10

15

## CLAIMS

1. A DNA synthesis reaction-enhancer comprising at least one kind selected from the group consisting of acidic substances and cationic complexes.

5

2. The DNA synthesis reaction-enhancer according to claim 1, wherein said acidic substance is an acidic macromolecular substance.

10

3. The DNA synthesis reaction-enhancer according to claim 2, wherein said acidic macromolecular substance is an acidic polysaccharide.

15

4. The DNA synthesis reaction-enhancer according to claim 2 or 3, wherein said acidic macromolecular substance is one or more substances selected from the group consisting of sulfated-fucose-containing polysaccharides, dextran sulfate, carrageenan, heparin, rhamnam sulfate, dermatan sulfate (chondroitin sulfate B), heparan sulfate, hyaluronic acid, alginic acid, pectin, polyglutamic acids, polyacrylic acids, polyvinyl sulfates, polystyrene sulfates, carrageenan, DNA and salts thereof.

20

5. The DNA synthesis reaction-enhancer according to claim 4, wherein said sulfated-fucose-containing polysaccharide is sulfated-fucose-containing polysaccharide-F or sulfated-fucose-containing polysaccharide-U.

25

6. The DNA synthesis reaction-enhancer according to claim 1, wherein said cationic complex is a transition metal complex.

7. The DNA synthesis reaction-enhancer according to claim 6, wherein a central atom in said transition metal complex is a transition element of the Group VIII of the elemental periodic table.

5

8. The DNA synthesis reaction-enhancer according to claim 7, wherein said transition element is one or more elements selected from the group consisting of cobalt, rhodium and iridium.

10

9. The DNA synthesis reaction-enhancer according to claim 8, wherein said transition metal complex is one or more kinds selected from the group consisting of  $[\text{Co}(\text{NH}_3)_6]\text{Cl}_3$ ,  $[\text{Co}(\text{H}_2\text{NCH}_2\text{CH}_2\text{NH}_2)_3]\text{Cl}_3$  and  $[\text{Rh}(\text{H}_2\text{NCH}_2\text{CH}_2\text{NH}_2)_3]\text{Cl}_3$ .

15

10. A DNA synthesis method characterized in that during a DNA synthesis reaction a reaction is carried out in the presence of the DNA synthesis reaction-enhancer of any one of claims 1 to 9 by using DNA polymerase.

11. The DNA synthesis method according to claim 10, wherein two or more kinds of DNA polymerases are used.

20

12. The DNA synthesis method according to claim 11, wherein one DNA polymerase having  $3' \rightarrow 5'$  exonuclease activity, and the other DNA polymerase having no  $3' \rightarrow 5'$  exonuclease activity are used.

25

13. The DNA synthesis method according to claim 11, wherein two or more

kinds of DNA polymerases each having  $3' \rightarrow 5'$  exonuclease activity are used.

14. The DNA synthesis method according to claim 13, wherein  $\alpha$ -type DNA polymerase and non- $\alpha$ , non-polI type DNA polymerase are used.

5

15. The DNA synthesis method according to any one of claims 10 to 14, which is carried out by polymerase chain reaction (PCR) method.

10

16. A DNA synthesis reaction composition comprising the DNA synthesis reaction-enhancer of any one of claims 1 to 9.

17. The DNA synthesis reaction composition according to claim 16, further comprising DNA polymerase.

15

18. The DNA synthesis reaction composition according to claim 17, wherein the composition comprises two or more kinds of DNA polymerases.

20

19. The DNA synthesis reaction composition according to claim 18, wherein the composition comprises two or more kinds of DNA polymerases each having  $3' \rightarrow 5'$  exonuclease activity.

20. The DNA synthesis reaction composition according to claim 19, wherein the composition comprises  $\alpha$ -type DNA polymerase and non- $\alpha$ , non-polI type DNA polymerase.

25

21. The DNA synthesis reaction composition according to claim 18, wherein the composition comprises one DNA polymerase having 3' → 5' exonuclease activity, and the other DNA polymerase having no 3' → 5' exonuclease activity.

5 22. A DNA synthesis reaction composition comprising two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity.

10 23. The DNA synthesis reaction composition according to claim 22, wherein the composition comprises  $\alpha$ -type DNA polymerase and non- $\alpha$ , non-poll type DNA polymerase.

15 24. A DNA synthesis method characterized in that during a DNA synthesis reaction two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity are used.

25 25. The DNA synthesis method according to claim 24, wherein  $\alpha$ -type DNA polymerase and non- $\alpha$ , non-poll type DNA polymerase are used.

20 26. The DNA synthesis method according to claim 24 or 25, which is carried out by polymerase chain reaction (PCR) method.

27. A kit for use in *in vitro* DNA synthesis, comprising two or more kinds of DNA polymerases each having 3' → 5' exonuclease activity.

25 28. The kit according to claim 27, wherein the kit comprises  $\alpha$ -type DNA

polymerase and non- $\alpha$ , non-poll type DNA polymerase.

29. The kit according to claim 27 or 28, further comprising a reagent usable for DNA synthesis.

5

30. The kit according to any one of claims 27 to 29, wherein said DNA polymerase is a thermostable DNA polymerase.

31. A kit for use in *in vitro* DNA synthesis, wherein the kit comprises the DNA synthesis reaction-enhancer of any one of claims 1 to 9 and DNA polymerase.

10

32. The kit according to claim 31, further comprising a reagent usable for DNA synthesis.

15

33. The kit according to claim 31 or 32, which comprises two or more kinds of DNA polymerases.

34. The kit according to any one of claims 31 to 33, wherein said DNA polymerase is a thermostable DNA polymerase.

20



# BIRCH, STEWART, KOLASCH & BIRCH, LLP

P.O. Box 747 • Falls Church, Virginia 22040-0747

Telephone: (703) 205-8000 • Facsimile: (703) 205-8050

ATTORNEY DOCKET NO.  
1422-443P

PLEASE NOTE:  
YOU MUST  
COMPLETE THE  
FOLLOWING:



## COMBINED DECLARATION AND POWER OF ATTORNEY FOR PATENT AND DESIGN APPLICATIONS

As a below named inventor, I hereby declare that: my residence, post office address and citizenship are as stated next to my name; that I verily believe that I am the original, first and sole inventor (if only one inventor is named below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Insert Title: → METHOD FOR SYNTHESIZING DNA

Fill in Appropriate  
Information —  
For Use →  
Without  
Specification  
Attached:

the specification of which is attached hereto. If not attached hereto,

the specification was filed on \_\_\_\_\_ as  
United States Application Number \_\_\_\_\_;  
and amended on \_\_\_\_\_ (if applicable); and/or  
the specification was filed on April 21, 1999 as PCT  
International Application Number PCT/JP99/02121; and was  
amended under PCT Article 19 on \_\_\_\_\_ (if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I do not know and do not believe the same was ever known or used in the United States of America before my or our invention thereof, or patented or described in any printed publication in any country before my or our invention thereof or more than one year prior to this application, that the same was not in public use or on sale in the United States of America more than one year prior to this application, that the invention has not been patented or made the subject of an inventor's certificate issued before the date of this application in any country foreign to the United States of America on an application filed by me or my legal representatives or assigns more than twelve months (six months for designs) prior to this application, and that no application for patent or inventor's certificate on this invention has been filed in any country foreign to the United States of America prior to this application by me or my legal representatives or assigns, except as follows.

I hereby claim foreign priority benefits under Title 35, United States Code, §119 (a)-(d) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Insert Priority  
Information: →  
(if appropriate)

Prior Foreign Application(s)			Priority Claimed	
<u>10-114005</u>	<u>Japan</u>	<u>April 23, 1998</u>	<input checked="" type="checkbox"/> Yes	<input type="checkbox"/> No
(Number)	(Country)	(Month / Day / Year Filed)		
<u>10-315243</u>	<u>Japan</u>	<u>November 6, 1998</u>	<input checked="" type="checkbox"/> Yes	<input type="checkbox"/> No
(Number)	(Country)	(Month / Day / Year Filed)		
_____	_____	_____	<input type="checkbox"/> Yes	<input type="checkbox"/> No
(Number)	(Country)	(Month / Day / Year Filed)		
_____	_____	_____	<input type="checkbox"/> Yes	<input type="checkbox"/> No
(Number)	(Country)	(Month / Day / Year Filed)		

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below.

Insert Provisional  
Application(s): →  
(if any)

(Application Number)	(Filing Date)
_____	_____
(Application Number)	(Filing Date)

All Foreign Applications, if any, for any Patent or Inventor's Certificate Filed More than 12 Months (6 Months for Designs) Prior to the Filing Date of This Application:

Insert Requested  
Information: →  
(if appropriate)

Country	Application Number	Date of Filing (Month / Day / Year)
_____	_____	_____
_____	_____	_____

I hereby claim the benefit under Title 35, United States Code, §120 of any United States and/or PCT application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States and/or PCT application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Insert Prior U.S.  
Application(s): →  
(if any)

(Application Number)	(Filing Date)	(Status — patented, pending, abandoned)
_____	_____	_____
(Application Number)	(Filing Date)	(Status — patented, pending, abandoned)

I hereby appoint the following attorneys to prosecute this application and/or an international application based on this application and to transact all business in the Patent and Trademark Office connected therewith and in connection with the resulting patent based on instructions received from the entity who first sent the application papers to the attorneys identified below, unless the inventor(s) or assignee provides said attorneys with a written notice to the contrary:

Raymond C. Stewart	(Reg. No. 21,066)	Terrell C. Birch	(Reg. No. 19,382)
Joseph A. Kolasch	(Reg. No. 22,463)	James M. Slattery	(Reg. No. 28,380)
Bernard L. Sweeney	(Reg. No. 24,448)	Michael K. Mutter	(Reg. No. 29,680)
Charles Gorenstein	(Reg. No. 29,271)	Gerald M. Murphy, Jr.	(Reg. No. 28,977)
Leonard R. Svensson	(Reg. No. 30,330)	Terry L. Clark	(Reg. No. 32,644)
Andrew D. Meikle	(Reg. No. 32,868)	Marc S. Weiner	(Reg. No. 32,181)
Joe McKinney Muncy	(Reg. No. 32,334)	C. Joseph Faraci	(Reg. No. 32,350)
Donald J. Daley	(Reg. No. 34,313)	John W. Bailey	(Reg. No. 32,881)
John A. Castellano	(Reg. No. 35,094)		

Send Correspondence to: **BIRCH, STEWART, KOLASCH & BIRCH, LLP**  
P.O. Box 747 • Falls Church, Virginia 22040-0747  
Telephone: (703) 205-8000 • Facsimile: (703) 205-8050

**PLEASE NOTE:  
YOU MUST  
COMPLETE  
THE  
FOLLOWING:**



I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of First or  
Sole Inventor:  
Insert Name of  
Inventor  
Insert Date This  
Document is Signed

Insert Residence  
Insert Citizenship →

Insert Post Office  
Address →

Full Name of Second  
Inventor, if any:  
see above

Full Name of Third  
Inventor, if any:  
see above

Full Name of Fourth  
Inventor, if any:  
see above

Full Name of Fifth  
Inventor, if any:  
see above

<b>GIVEN NAME</b> Kiyozo	<b>FAMILY NAME</b> ASADA	<b>INVENTOR'S SIGNATURE</b> <i>Kiyozo Asada</i>	<b>DATE*</b> 2000.9.19
<b>Residence (City, State &amp; Country)</b> Koga-gun, Shiga 520-3333 Japan		<b>CITIZENSHIP</b> Japan	
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b> 3-20-9, Kibogaoka, Konan-cho, Koga-gun, Shiga 520-3333 Japan			
<b>GIVEN NAME</b> Takashi	<b>FAMILY NAME</b> UEMORI	<b>INVENTOR'S SIGNATURE</b> <i>Takashi Uemori</i>	<b>DATE*</b> 2000.9.19
<b>Residence (City, State &amp; Country)</b> Otsu-shi, Shiga 520-2141 Japan		<b>CITIZENSHIP</b> Japan	
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b> 1-16-709, Oe 3-chome, Otsu-shi, Shiga 520-2141 Japan			
<b>GIVEN NAME</b> Yoshimi	<b>FAMILY NAME</b> SATO	<b>INVENTOR'S SIGNATURE</b> <i>Yoshimi Sato</i>	<b>DATE*</b> 2000.10.2
<b>Residence (City, State &amp; Country)</b> Kurita-gun, Shiga 520-3031 Japan		<b>CITIZENSHIP</b> Japan	
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b> 390-7-C-1010, Oaza Heso, Ritto-cho, Kurita-gun, Shiga 520-3031 Japan			
<b>GIVEN NAME</b> Tomoko	<b>FAMILY NAME</b> FUJITA	<b>INVENTOR'S SIGNATURE</b> <i>Tomoko Fujita</i>	<b>DATE*</b> 2000.9.28
<b>Residence (City, State &amp; Country)</b> Takatsuki-shi, Osaka 569-1144 Japan		<b>CITIZENSHIP</b> Japan	
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b> 25-10, Ohata-cho, Takatsuki-shi, Osaka 569-1144 Japan			
<b>GIVEN NAME</b> Kazue	<b>FAMILY NAME</b> MIYAKE	<b>INVENTOR'S SIGNATURE</b> <i>Kazue Miyake</i>	<b>DATE*</b> 2000.9.22
<b>Residence (City, State &amp; Country)</b> Uji-shi, Kyoto 611-0011 Japan		<b>CITIZENSHIP</b> Japan	
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b> 34-7, Sanbanwari, Gokanosyo, Uji-shi, Kyoto 611-0011 Japan			

I hereby appoint the following attorneys to prosecute this application and/or an international application based on this application and to transact all business in the Patent and Trademark Office connected therewith and in connection with the resulting patent based on instructions received from the entity who first sent the application papers to the attorneys identified below, unless the inventor(s) or assignee provides said attorneys with a written notice to the contrary:

Raymond C. Stewart (Reg. No. 21,066)  
 Joseph A. Kolasch (Reg. No. 22,463)  
 Bernard L. Sweeney (Reg. No. 24,448)  
 Charles Gorenstein (Reg. No. 29,271)  
 Leonard R. Svensson (Reg. No. 30,330)  
 Andrew D. Meikle (Reg. No. 32,868)  
 Joe McKinney Muncy (Reg. No. 32,334)  
 Donald J. Daley (Reg. No. 34,313)  
 John A. Castellano (Reg. No. 35,094)

Terrell C. Birch (Reg. No. 19,382)  
 James M. Slattery (Reg. No. 28,380)  
 Michael K. Mutter (Reg. No. 29,680)  
 Gerald M. Murphy, Jr. (Reg. No. 28,977)  
 Terry L. Clark (Reg. No. 32,644)  
 Marc S. Weiner (Reg. No. 32,181)  
 C. Joseph Faraci (Reg. No. 32,350)  
 John W. Bailey (Reg. No. 32,881)

Send Correspondence to: **BIRCH, STEWART, KOLASCH & BIRCH, LLP**  
 P.O. Box 747 • Falls Church, Virginia 22040-0747  
 Telephone: (703) 205-8000 • Facsimile: (703) 205-8050

PLEASE NOTE:  
 YOU MUST  
 COMPLETE  
 THE  
 FOLLOWING:

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of First or  
 Sole Inventor:  
 Insert Name of  
 Inventor  
 Insert Date This  
 Document is Signed

Insert Residence  
 Insert Citizenship

Insert Post Office  
 Address

Full Name of Second  
 Inventor, if any:  
 see above

Full Name of Third  
 Inventor, if any:  
 see above

Full Name of Fourth  
 Inventor, if any:  
 see above

Full Name of Fifth  
 Inventor, if any:  
 see above

<b>GIVEN NAME</b> Osamu	<b>FAMILY NAME</b> TAKEDA	<b>INVENTOR'S SIGNATURE</b> Osamu Takeda	<b>DATE*</b> 2000.9.21
<b>Residence (City, State &amp; Country)</b> Hikone-shi, Shiga 521-1124 Japan			<b>CITIZENSHIP</b> Japan
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b> 340-1-811, Norada-cho, Hikone-shi, Shiga 521-1124 Japan			
<b>GIVEN NAME</b> Hiroyuki	<b>FAMILY NAME</b> MUKAI	<b>INVENTOR'S SIGNATURE</b> Hiroyuki Mukai	<b>DATE*</b> 2000.9.21
<b>Residence (City, State &amp; Country)</b> Moriyama-shi, Shiga 524-0102 Japan			<b>CITIZENSHIP</b> Japan
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b> 1461-82, Aza Minamikawa, Mizuho-cho, Moriyama-shi, Shiga 524-0102 Japan			
<b>GIVEN NAME</b> Ikunoshin	<b>FAMILY NAME</b> KATO	<b>INVENTOR'S SIGNATURE</b> Ikunoshin Kato	<b>DATE*</b> 2000.9.21
<b>Residence (City, State &amp; Country)</b> Uji-shi, Kyoto 611-0028 Japan			<b>CITIZENSHIP</b> Japan
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b> 1-1-150, Nanryo-cho, Uji-shi, Kyoto 611-0028 Japan			
<b>GIVEN NAME</b>	<b>FAMILY NAME</b>	<b>INVENTOR'S SIGNATURE</b>	<b>DATE*</b>
<b>Residence (City, State &amp; Country)</b>			<b>CITIZENSHIP</b>
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b>			
<b>GIVEN NAME</b>	<b>FAMILY NAME</b>	<b>INVENTOR'S SIGNATURE</b>	<b>DATE*</b>
<b>Residence (City, State &amp; Country)</b>			<b>CITIZENSHIP</b>
<b>POST OFFICE ADDRESS (Complete Street Address including City, State &amp; Country)</b>			

## SEQUENCE LISTING

<110> Takara Shuzo Co., Ltd.

<120> A method for DNA synthesis

<130> 99-017-PCT

<150> JP 10-114005

<151> 1998-4-23

<150> JP 10-315243

<151> 1998-11-6

<160> 18

<210> 1

<211> 23

<212> DNA

<213> Artificial Sequence

<400> 1

gatgagttcg tgtccgtaca act

23

<210> 2

<211> 22

<212> DNA

<213> Artificial Sequence

<400> 2

acaaagccag ccggaatata tg

22

<210> 3

<211> 35

<212> DNA

<213> Artificial Sequence

<400> 3

gatgagttcg tgtccgtaca actggcgtaa tcatg

35

<210> 4

<211> 25

<212> DNA

<213> Artificial Sequence

<400> 4

ggttatcgaa atcagccaca gcgcc

25

<210> 5

<211> 23

<212> DNA

<213> Artificial Sequence

<400> 5

gcgtaccttt gtctcaggg caa

23

<210> 6  
 <211> 22  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 6  
 gatagctgtc gtcataggac tc 22

<210> 7  
 <211> 23  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 7  
 cttaaccagt gcgctgagtg act 23

<210> 8  
 <211> 28  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 8  
 ttgccacttc cgtcaaccag gcttatca 28

<210> 9  
 <211> 29  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 9  
 tgtccgtcag ctcataacgg tacttcacg 29

<210> 10  
 <211> 28  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 10  
 atatctggcg gtgcaatadc ggtactgt 28

<210> 11  
 <211> 28  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 11  
 gacaatctgg aatacgccac ctgacttg 28

<210> 12  
 <211> 36  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 12  
 gggcggcgac ctgcggggtt ttgctattt atgaaa 36

<210> 13  
 <211> 36  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 13  
 taacctgtcg gatcaccgga aaggaccgt aaagtg 36

<210> 14  
 <211> 35  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 14  
 ggtggcgtg caaatgcaat ctctgtgcc ccaac 35

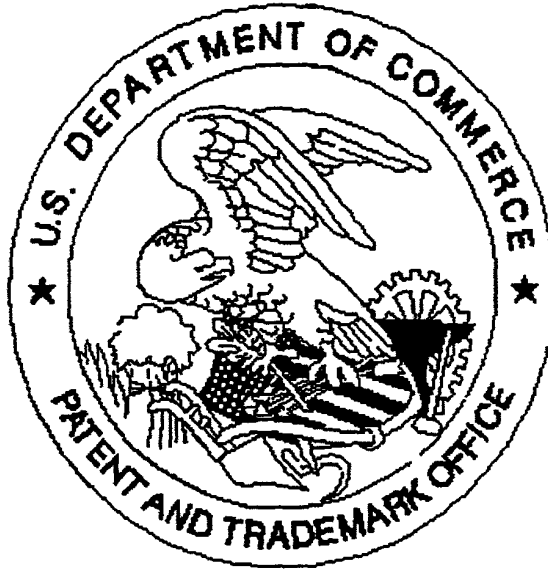
<210> 15  
 <211> 35  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 15  
 ttatgtatgc cgcgtatcag ctctatgtct ggctc 35

<210> 16  
 <211> 35  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 16  
 atcatctaac ctgttctgga aaacgcttgc gcagc 35

<210> 17  
 <211> 19  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 17  
 aagcgctgg cagtgtacc 19

<210> 18  
 <211> 21  
 <212> DNA  
 <213> Artificial Sequence  
 <400> 18  
 ctctggcgtt cagtattgt c 21

United States Patent & Trademark Office  
Office of Initial Patent Examination -- Scanning Division



Application deficiencies found during scanning:

☒ Page(s) 0 of Drawings were not present  
for scanning. (Document title)

☐ Page(s) \_\_\_\_\_ of \_\_\_\_\_ were not present  
for scanning. (Document title)

☐ *Scanned copy is best available.*